

# Genetic mapping of Sclerotinia basal stalk rot resistance and oxalic acid tolerance in two sunflower recombinant inbred line populations

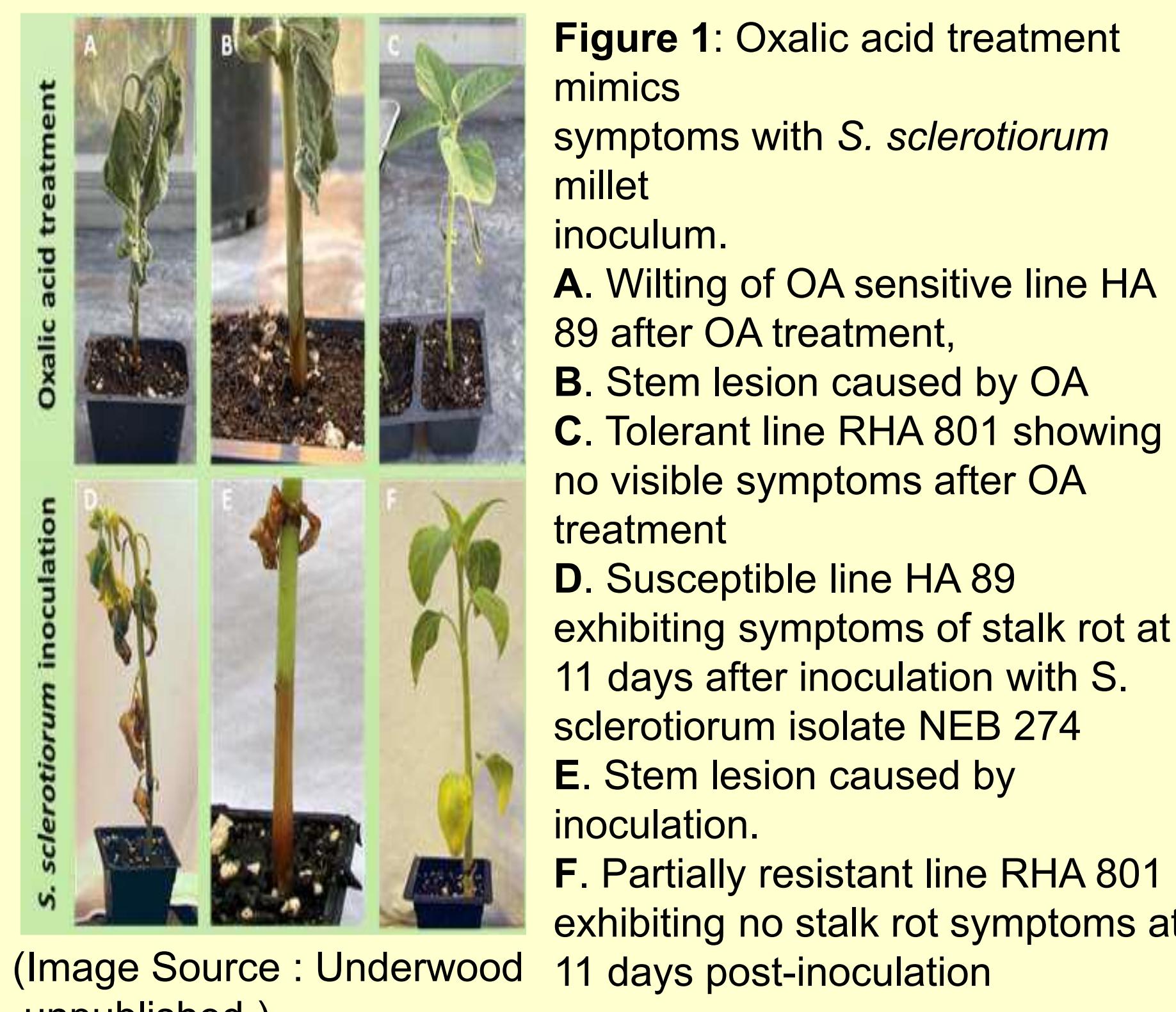
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## Research Background

- Sclerotinia basal stalk rot (BSR) of sunflower (*Helianthus annuus L.*) is a destructive disease incited by the necrotrophic fungal pathogen *Sclerotinia sclerotiorum*, which causes significant yield losses worldwide.
- Oxalic acid (OA) is a key virulence factor in *Sclerotinia sclerotiorum* pathogenesis inducing plant cell death, creating an acidic environment, aiding fungal penetration, and compromising host cell walls.
- Typical symptoms of BSR include- wilting in aerial tissues and basal stem lesions
- Noyes and Hancock identified OA as a wilt-inducing toxin from *Sclerotinia*-infected sunflower tissues
- Previously, while checking the oxalic acid tolerance in several stalk rot-susceptible and resistant lines, the roots of the plants were treated by soil drench with 60 mM potassium oxalate (pH 5.6). This treatment demonstrated that symptoms of OA mirror BSR symptoms, suggesting that OA may play a role in disease development (Figure 1).



- Previous research has identified OA tolerance in the sunflower inbred lines HA 61 and RHA 801, which also exhibit partial resistance to BSR (Figure 2).

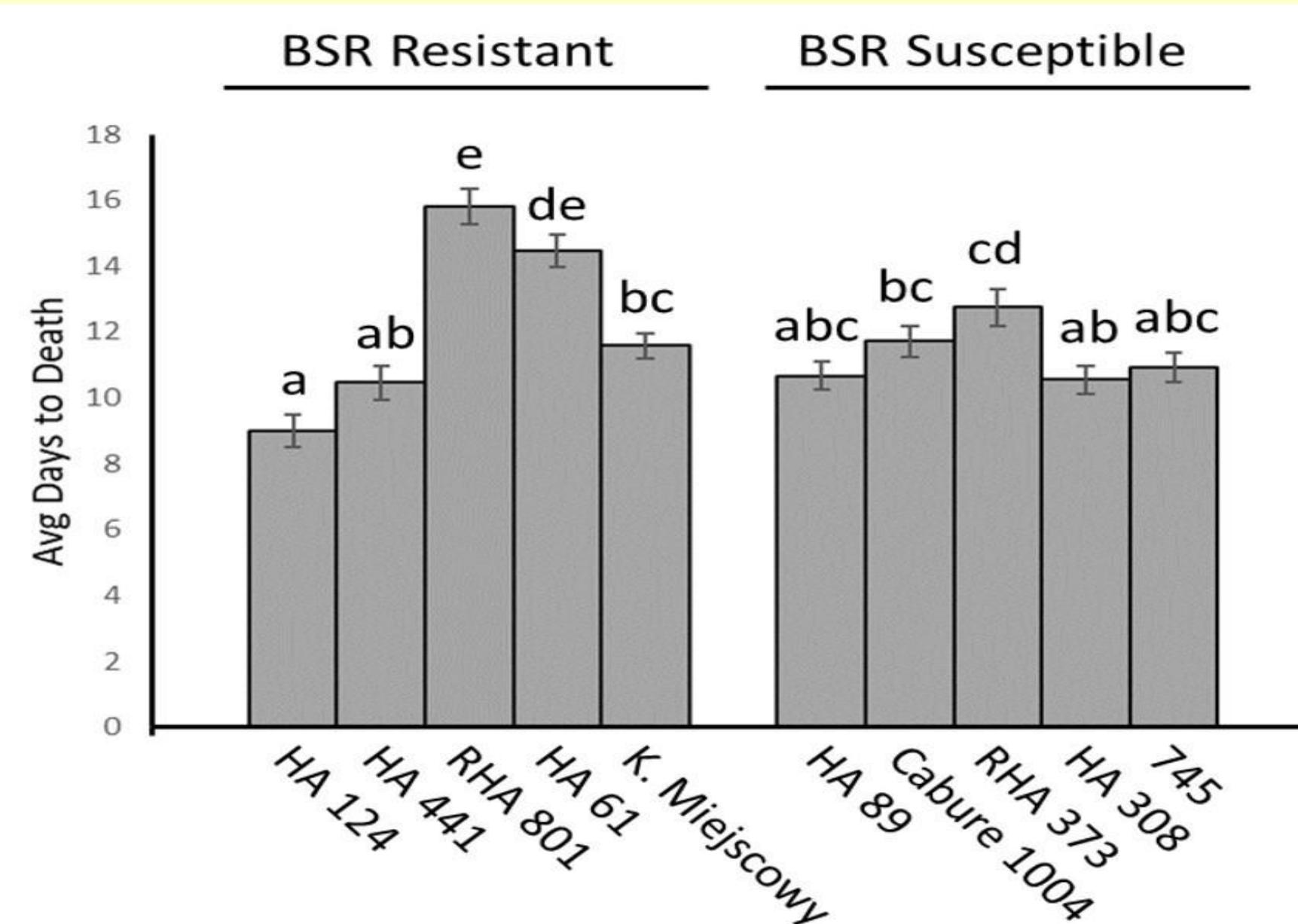


Figure 2: Evaluation of sunflower genotypes with OA

- The degree to which OA tolerance contributes to BSR resistance in HA 61 and RHA 801, if at all, is unknown.
- Therefore, we developed recombinant inbred line populations to facilitate genetic characterization of the BSR resistance and OA tolerance traits in HA 61 and RHA 801.

## Rationale

- Understanding the relationship between BSR resistance and OA tolerance and the genetic loci underlying these traits may allow sunflower breeders to improve resistance to this important disease by combining specific resistance mechanisms.

## Specific Objectives

- To map QTL for BSR resistance after inoculation with *S. sclerotiorum*.
- To map QTL for OA tolerance using a soil drench assay.
- To assess the correlation between the two traits and determine if any mapped loci are overlapping.

## Preliminary Results

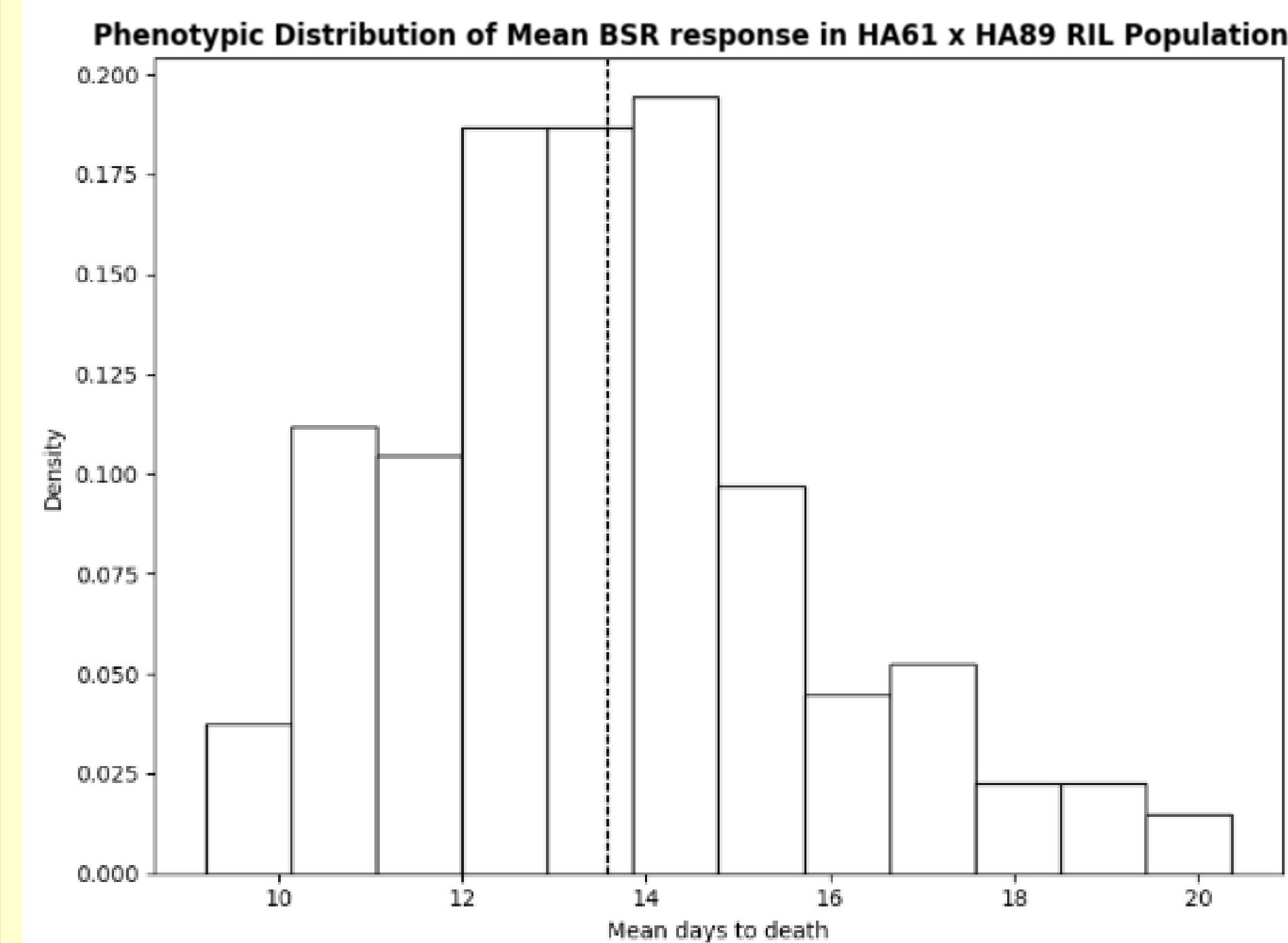


Figure 3 : Distribution of BSR response among RILs of the HA 61 × HA 89 population

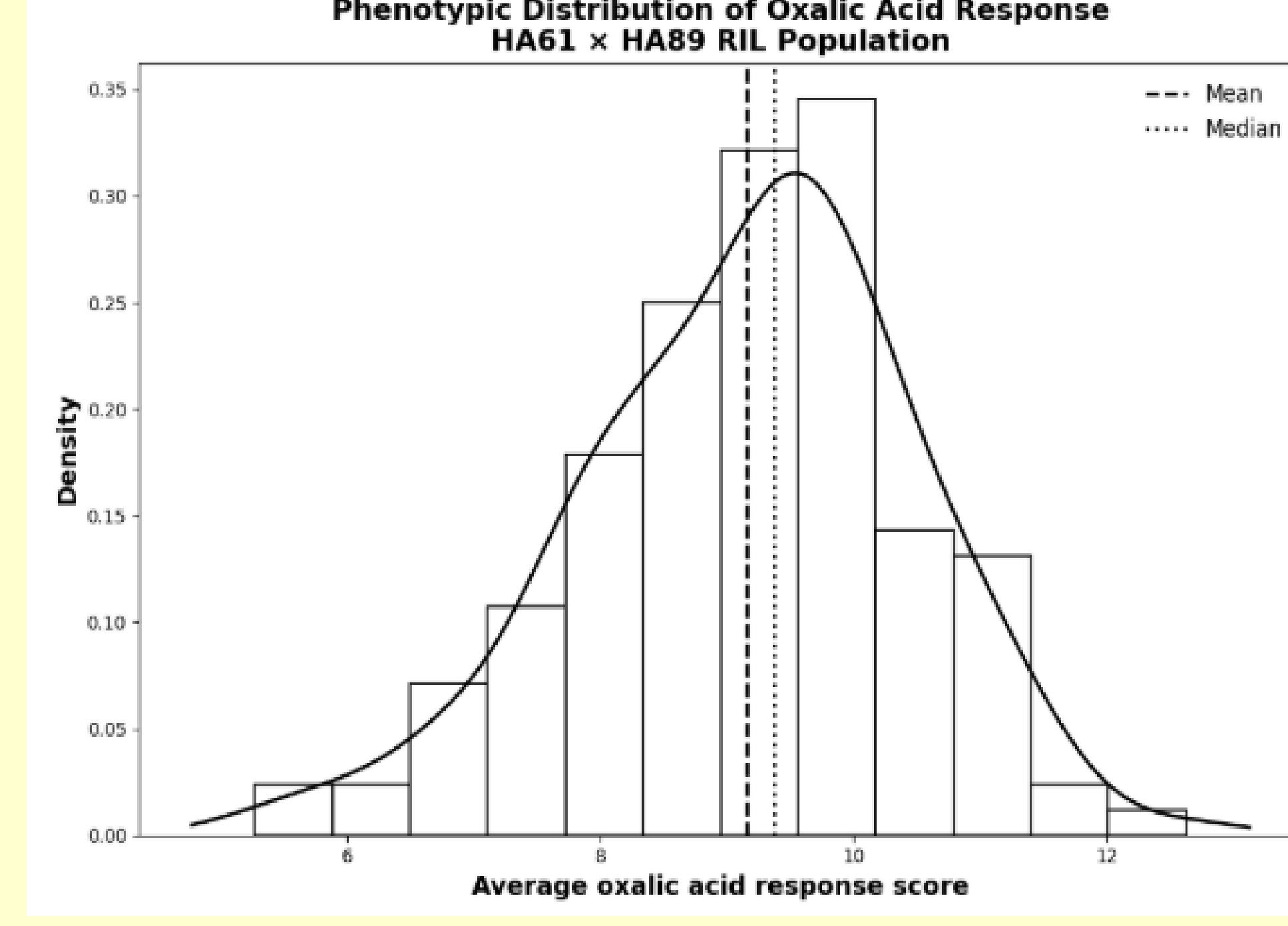


Figure 4: Distribution of OA response among RILs of the HA 61 × HA 89 population

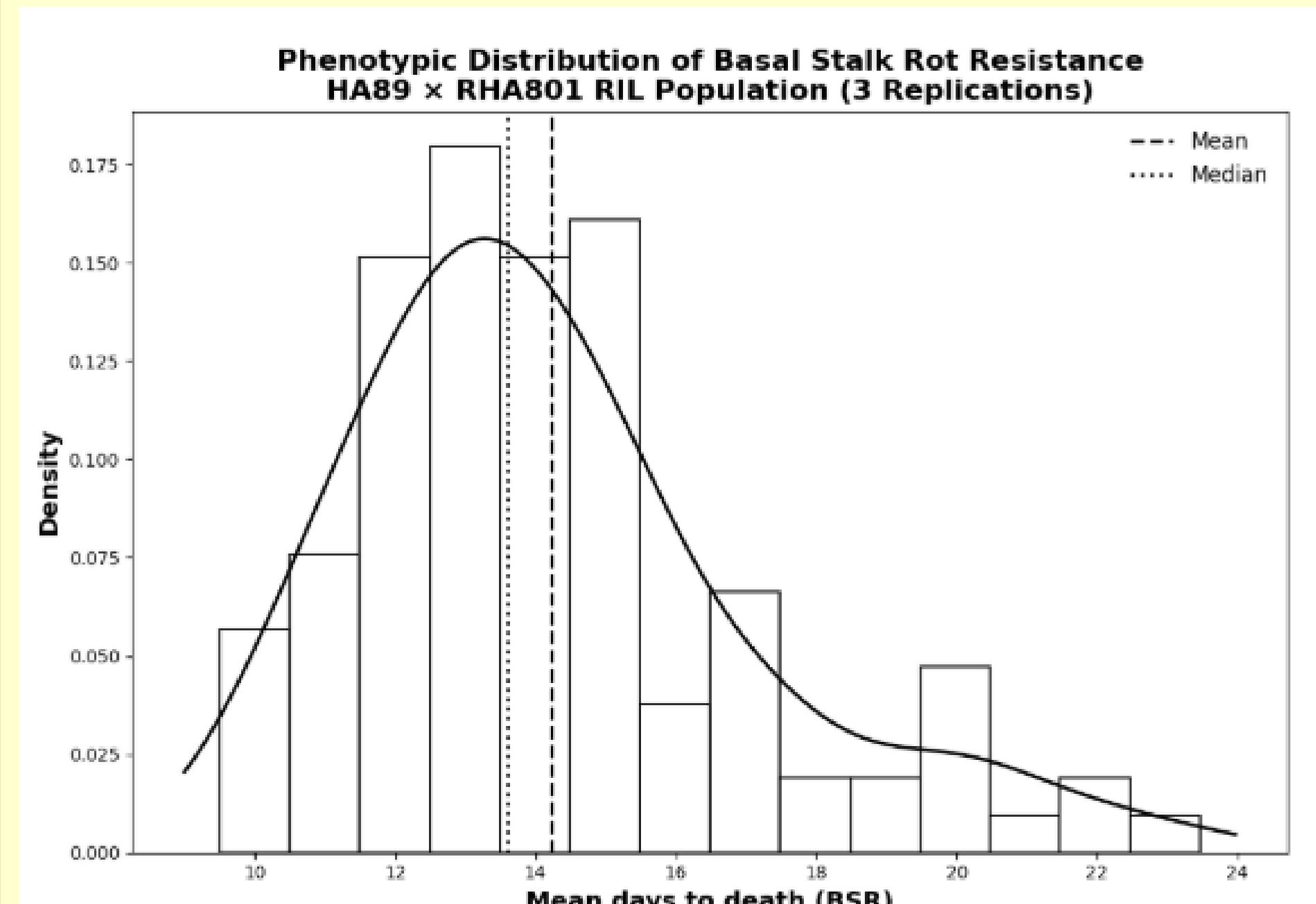


Figure 5 : Distribution of BSR response among RILs of the HA 89 x RHA 801 population

## Summary

- In conclusion, previous studies have identified inbred line HA 61 and RHA801 as displaying partial resistance to BSR in field trials and broad-spectrum resistance against diverse *S. sclerotiorum* isolates when evaluated in the greenhouse.
- Preliminary results for BSR phenotyping of the HA 61 x HA 89 and HA 89 x RHA 801 indicated the expected transgressive segregation consistent with quantitative resistance to BSR.
- Additionally, a single replication for Oxalic Acid response of HA 89 x RHA 801 RIL populations has been completed
- The results of this study will provide valuable insights into the genetic mechanisms governing resistance to BSR and the specific genes responsible for OA tolerance.

## Future Work

- In the future, our priority will be to complete OA response of HA 89 x RHA 801 RIL populations
- Sequence genetic samples and construct high-density linkage maps to enable robust QTL mapping.
- We aim to identify genomic regions associated with oxalic acid tolerance and BSR resistance while identifying overlapping QTLs to understand host-pathogen interactions.

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## Methodology

