



Breeding Pool and Market Class Differentiation in Leaf Physiology and Secondary Metabolism in Cultivated Sunflower

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Jordan Dowell - PhD 2021 (UCF)
USDA NIFA Postdoctoral Fellow (UC Davis)
GWAS + phenotypic data analysis



Alan Bowsher, PhD – HPLC phenotyping
Amna Jamshad, MD – leaf phenotyping
Rahul Shah, MD – leaf phenotyping
Lisa Donovan, PhD – phenotyping + design advice
John Burke, PhD – GWAS + interpretation advice

Why focus on leaves?

Leaf traits influence:

Assimilation (capture CO_2 + light)

Growth rate (product of leaf assimilation, leaf cost and lifespan, and plant allocation to leaves)

Abiotic stress tolerance (loss of water, nutrient resorption)

Biotic stress tolerance (herbivory + disease resistance)



Leaf Economic Spectrum



“Fast” Leaves

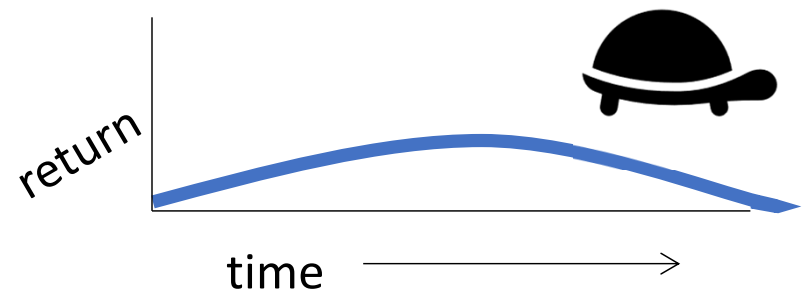
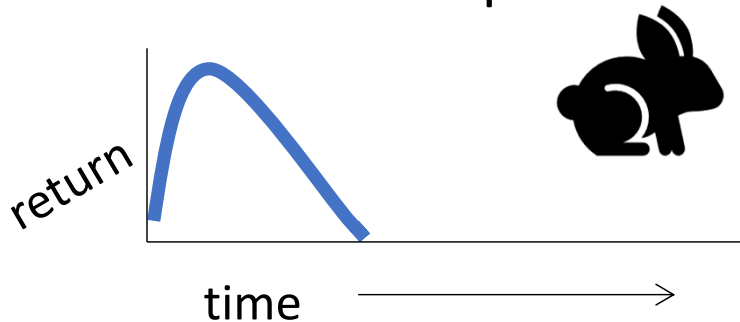
“Slow” Leaves

Resource acquisitive

High photosynthetic rate
High nutrient content
Low thickness/density
Short leaf lifespan

Resource conservative

Low photosynthetic rate
Low nutrient content
High thickness/density
Long leaf lifespan





Research Article

Inter- and intraspecific variation in leaf economic traits in wheat and maize

Adam R. Martin^{1,2*}, Christine E. Hale¹, Bruno E. L. Cerabolini³, Johannes H. C. Cornelissen⁴, Joseph Craine⁵, William A. Gough¹, Jens Kattge^{6,7} and

Functional Ecology

Functional Ecology 2017, 31, 604–612



doi: 10.1111/1365-2435.12790

Intraspecific trait variation across multiple scales: the leaf economics spectrum in coffee

Adam R. Martin^{1,2*}, Bruno Rapidel^{2,3}, Olivier Roupsard^{2,4}, Karel Van den Meersche^{2,4}, Eliza de Melo Virginia Filho², Mirza Barrios⁵ and Marnon E. Jones¹

Intraspecific variation in soy across the leaf economics spectrum

Fallon J. Hayes¹, Serra W. Buchanan¹, Brent Coleman², Andrew M. Gordon², Peter B. Reich^{3,4}, Naresh V. Thevathasan², Jan J. Wright⁵ and Adam R. Martin^{1,6,8*}

Journal of Experimental Botany, Vol. 69, No. 22 pp. 5599–5609, 2018
doi:10.1093/jxb/ery322 Advance Access publication 5 September 2018
This paper is available online free of all access charges (see http://jxb.oxfordjournals.org/open_access.html for further details)



RESEARCH PAPER

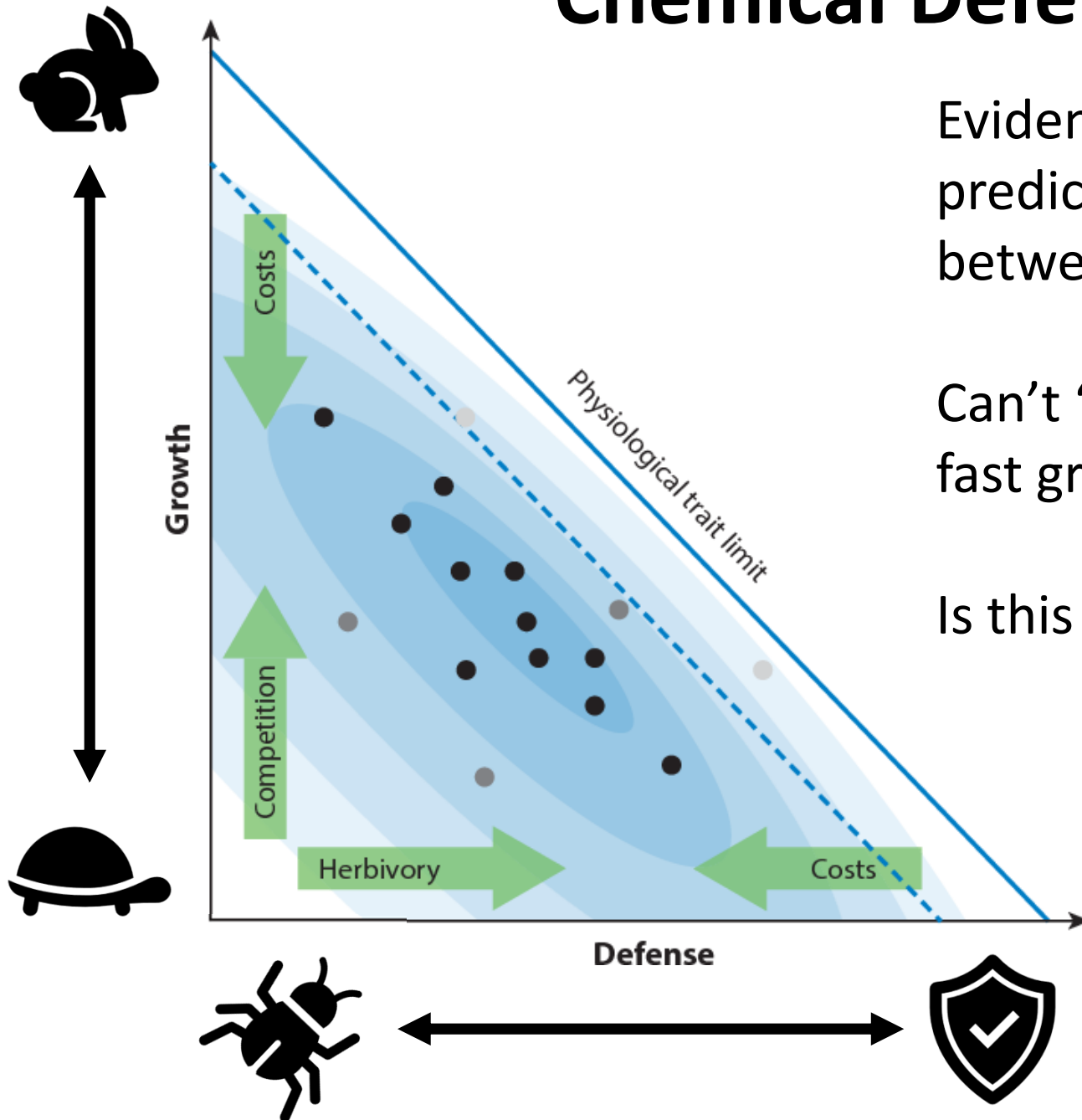
Leaf economics spectrum in rice: leaf anatomical, biochemical, and physiological trait trade-offs

Donliang Xiong^{1,2,*} and Jaime Flexas^{3,9}



Source: Kansas State
University Extension

Chemical Defense Investment



Evidence from wild plants predicts an inherent trade-off between growth and defense.

Can't "have it all" – don't see fast growth with high defense

Is this true within sunflower?

Market Classes + Breeding Groups

HA - maintainer

Oilseed

NonOil

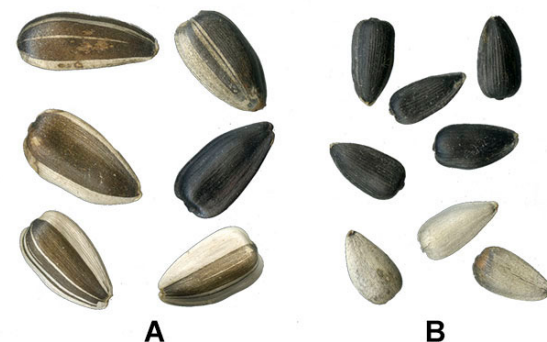
<i>HA Oil</i>	<i>HA NonOil</i>
<i>RHA Oil</i>	<i>RHA NonOil</i>
<i>Other Oil</i>	<i>Other NonOil</i>

RHA - restorer

**Others
(OPVs, etc)**

USDA Agricultural Marketing Service
U.S. DEPARTMENT OF AGRICULTURE

SUNFLOWER SEED



A. Confectionery Type. Black and white striped, thick-hulled, seed of the non-oil variety of sunflowers.

B. Oilseed Type. Black (some specialty varieties may be gray), thin-hulled, fairly large sized seed of the oil-producing variety of sunflower plant

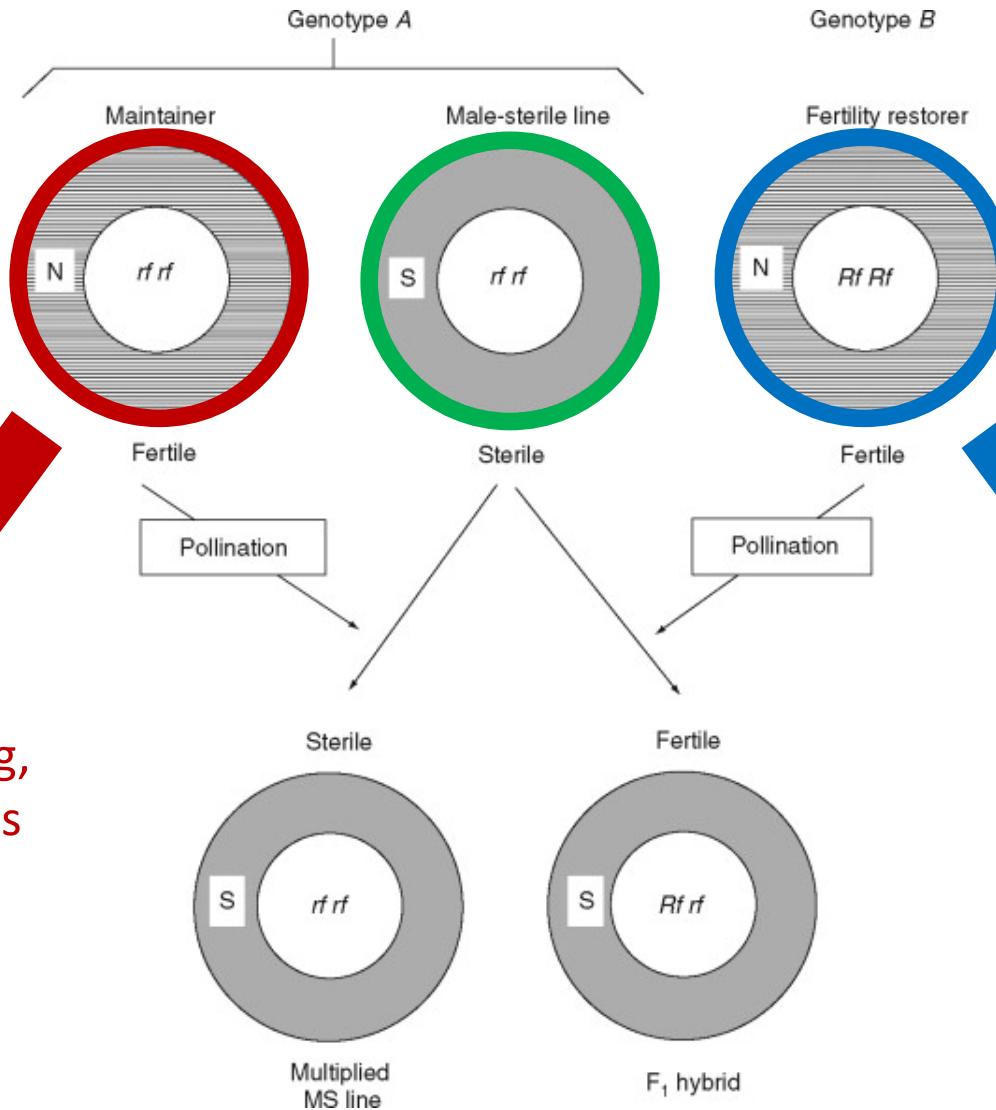


Male-sterile lines

HA - maintainer

*any germplasm without fertility-restoring alleles.

Propagated by selfing, can make inbred lines

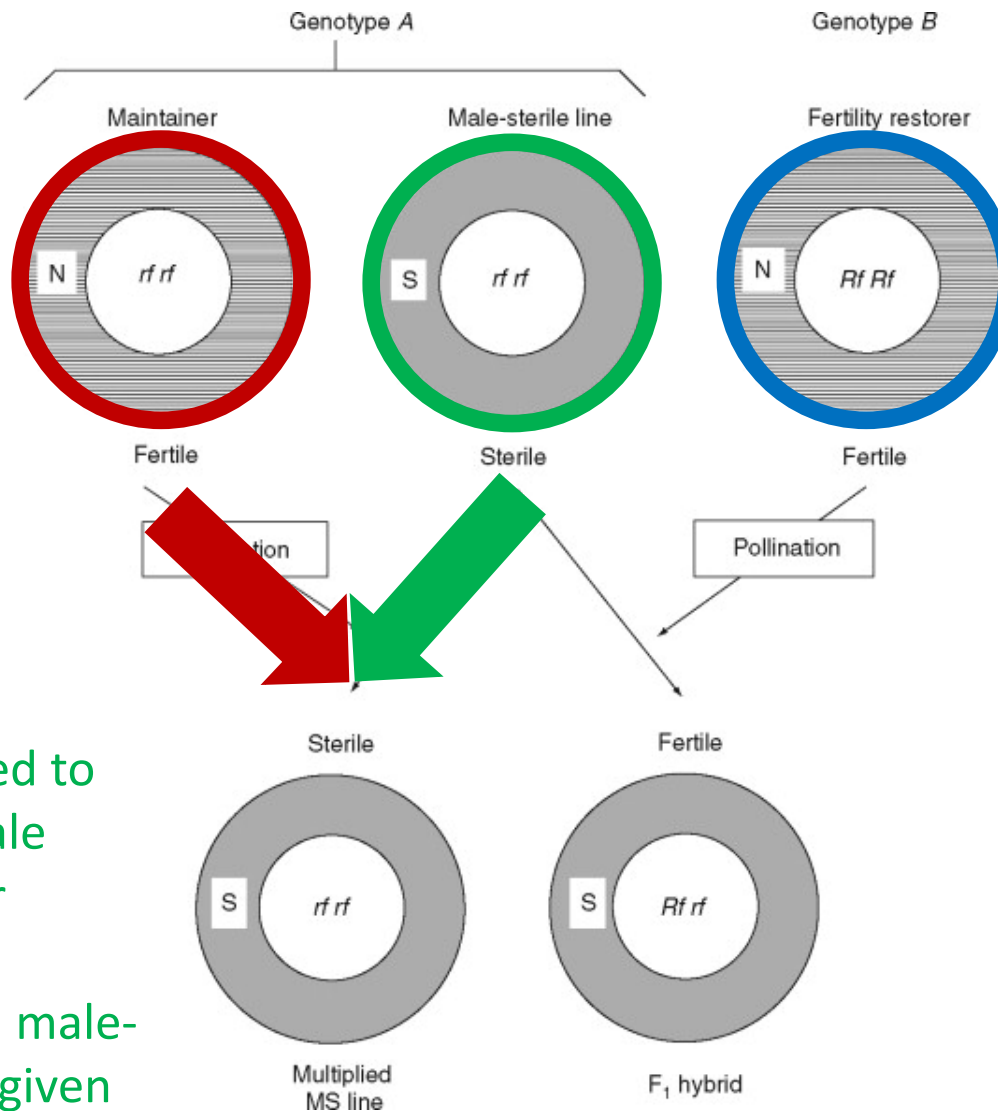


RHA - restorer

*only a few stable sources of nuclear restoration.

Propagated by selfing, can make inbred lines

Male-sterile lines



HA - maintainer

*any germplasm without fertility-restoring alleles.

RHA - restorer

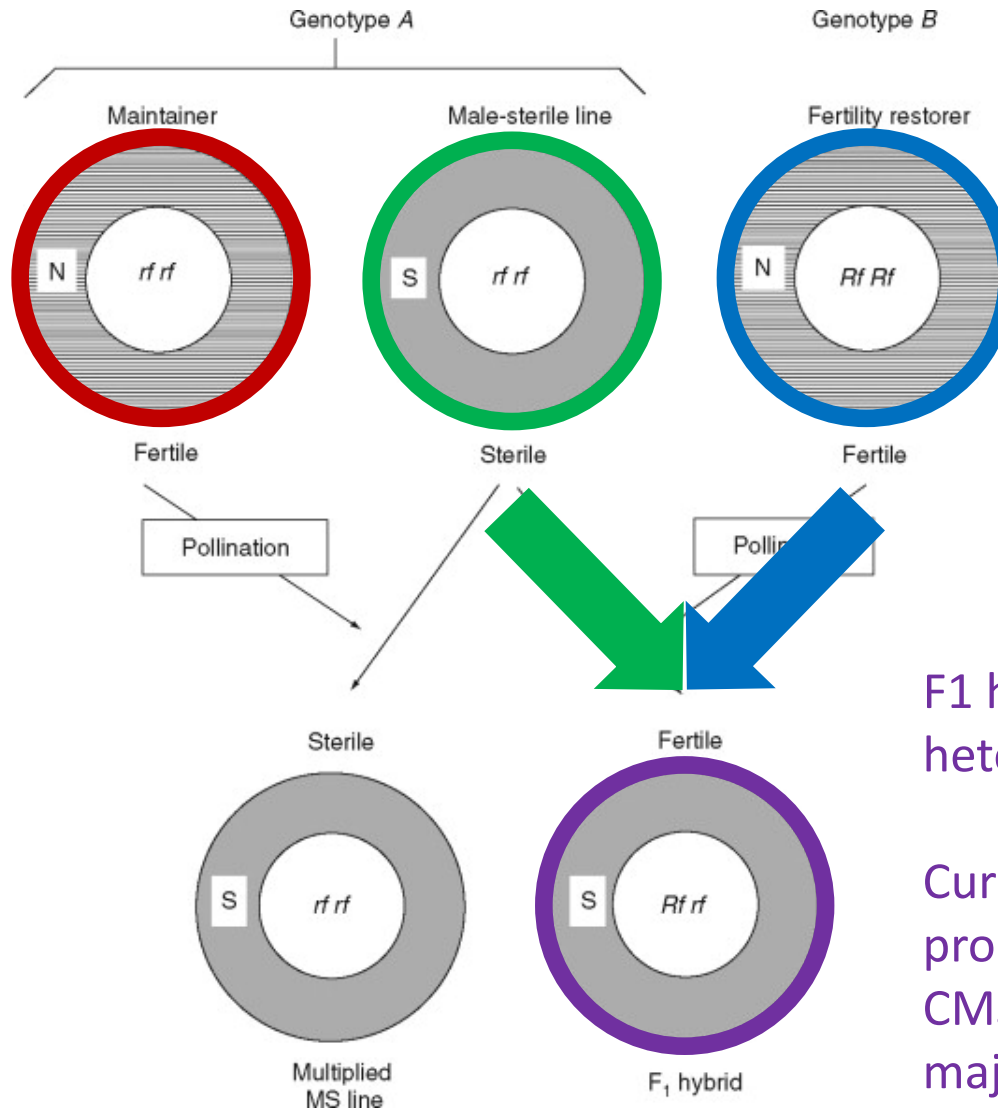
*only a few stable sources of nuclear restoration.

“Maintainers” are used to maintain the CMS-male sterile lines, and after several backcross generations you get a male-sterile version of any given “maintainer” genotype

Male-sterile lines

HA - maintainer

*any germplasm without fertility-restoring alleles.



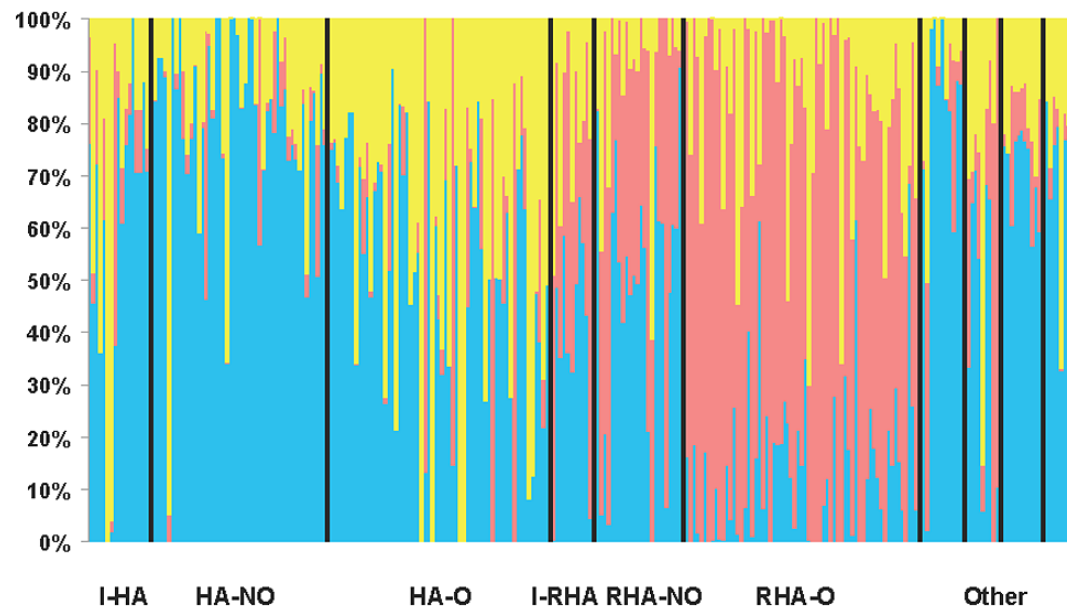
RHA - restorer

*only a few stable sources of nuclear restoration.

F1 hybrid seed with heterosis

Current commercial seed production mainly uses ONE CMS source (*PET1*) and ONE major fertility restoring gene – *Rf1*

Genetic Divergence in Breeding Groups/Market Classes



PLOS GENETICS

OPEN ACCESS PEER-REVIEWED

RESEARCH ARTICLE

2013

Association Mapping and the Genomic Consequences of Selection in Sunflower

Jennifer R. Mandel, Savithri Nambesan, John E. Bowers, Laura F. Marek, Daniel Ebert, Loren H. Rieseberg, Steven J. Knapp, John M. Burke

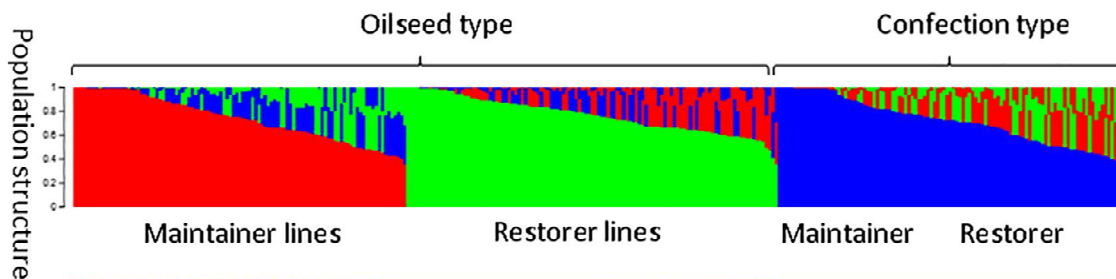
Published: March 21, 2013 • <https://doi.org/10.1371/journal.pgen.1003378>

frontiers
in Genetics

ORIGINAL RESEARCH
published: 14 March 2019
doi: 10.3389/fgene.2019.00216



2019



STRUCTURE – Bayesian clustering algorithm on 5000-9000 SNPs

Linkage Mapping and Genome-Wide Association Studies of the *Rf* Gene Cluster in Sunflower (*Helianthus annuus* L.) and Their Distribution in World Sunflower Collections

Zahirul I. Talukder¹, Guojia Ma¹, Brent S. Hulke², Chao-Chien Jan² and Lili Qi^{2*}

¹ Department of Plant Sciences, North Dakota State University, Fargo, ND, United States, ² Edward T. Schafer Agricultural Research Center, Agricultural Research Service, United States Department of Agriculture, Fargo, ND, United States

Today's Main Question

Given the known genetic divergence among breeding groups and market classes, have there been (unintended) phenotypic divergences?

Particular focus on:

leaf ecophysiology
leaf secondary metabolism
plant relative growth rate

Sunflower Association Mapping (SAM) Panel

288 cultivated lines representing an estimated 90% of allelic diversity within the USDA and INRA germplasm repositories.
Mean date of introduction = $1990 \pm 7\text{yr}$ (SD)

Multiple genetic maps, most recently full-genome resequencing data available for 261 lines, resulting in ~1.4 million SNP map (~3.6 Gb genome)

Handy “Core 12” set of lines representing ~half of allelic diversity within USDA + INRA germplasm



Theor Appl Genet (2011) 123:693–704
DOI 10.1007/s00122-011-1619-3

ORIGINAL PAPER

Genetic diversity and population structure in cultivated sunflower and a comparison to its wild progenitor, *Helianthus annuus* L

J. R. Mandel · J. M. Dechaine · L. F. Marek ·
J. M. Burke

2011

Leaf Ecophysiology + Secondary Metabolism

288 lines grown under high-resource conditions

Pest/pathogen-free environment
(constitutive ecophysiology and
metabolite production)

Sampling standardized by stage (V8-V10)

Pair of MRFELs taken for trait assessment
common ecophysiological traits
snap-frozen tissue analyzed by HPLC



Leaf Ecophysiology

PC1 – leaf economics (35%)

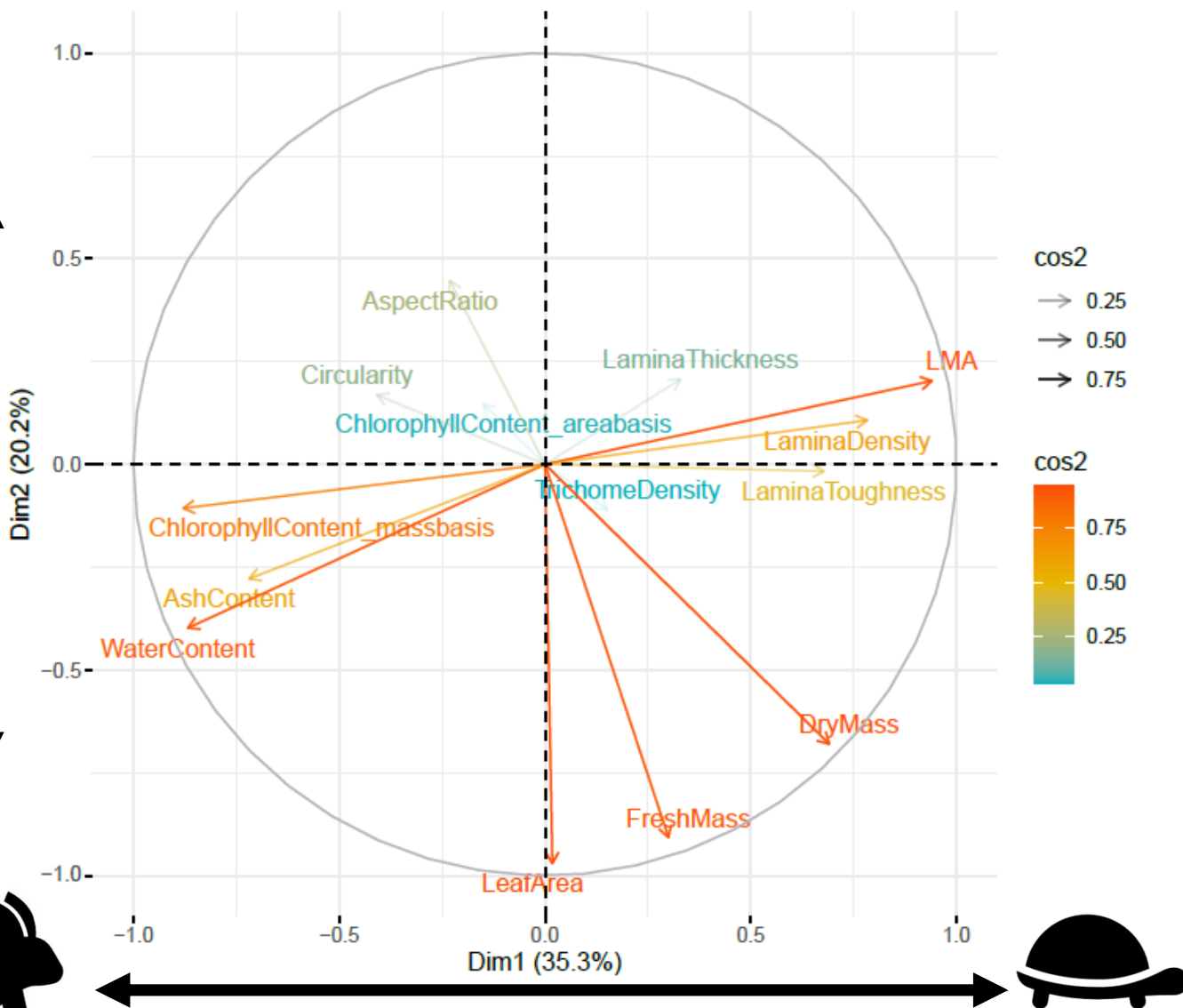
PC2 – leaf size (20%)



Small

Leaf Size

Large



Resource Acquisitive
"Fast"

Traditional
Leaf Economics Spectrum

Resource Conservative
"Slow"



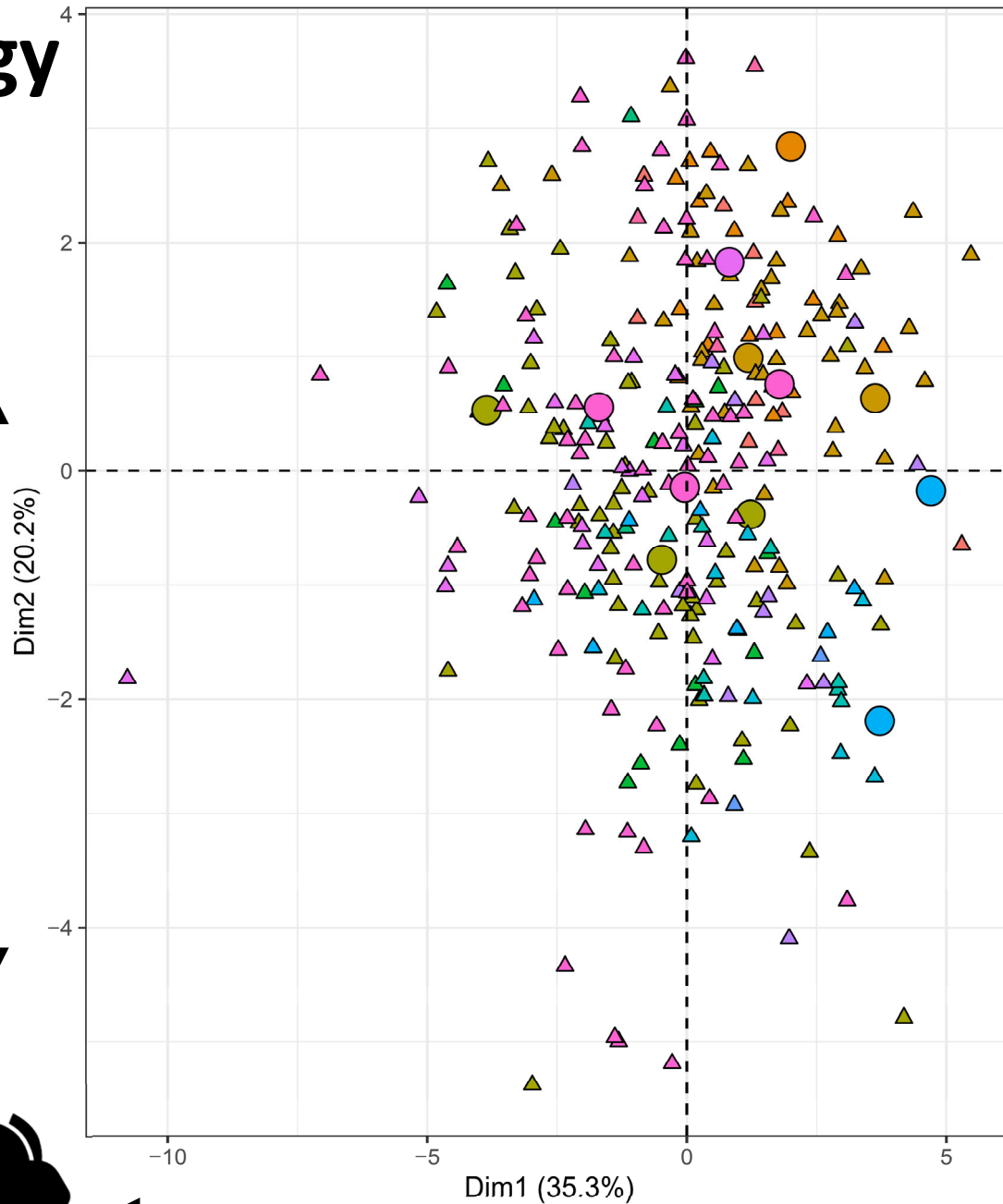
Leaf Ecophysiology

Interspersion of breeding groups and market classes, small differences

For leaf economics (PC1), RHA on average “faster” than HA, effect size $d=0.09$ (Bayes Factor=51)

Oilseed lines had larger leaves than NonOil, effect size $d=0.13$ (BF=31)

Small
Leaf Size
Large



Resource Acquisitive
“Fast”

Traditional
Leaf Economics Spectrum

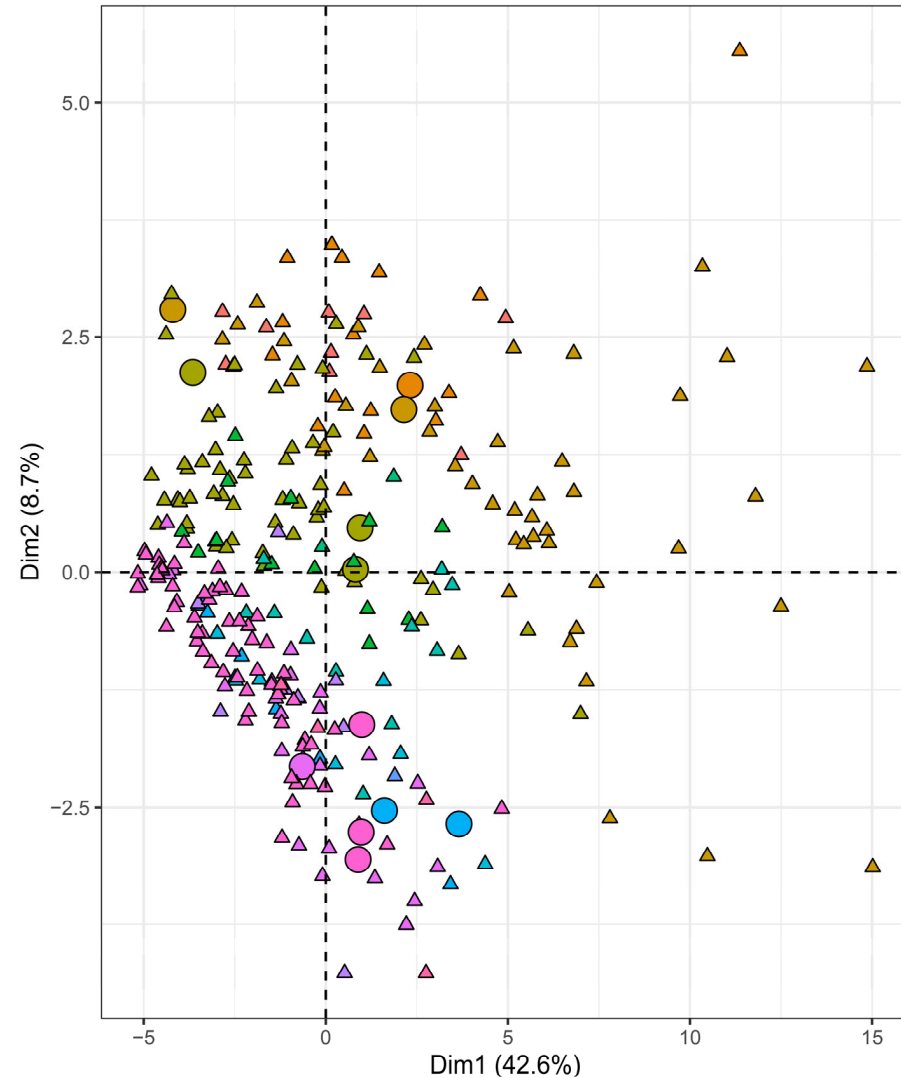
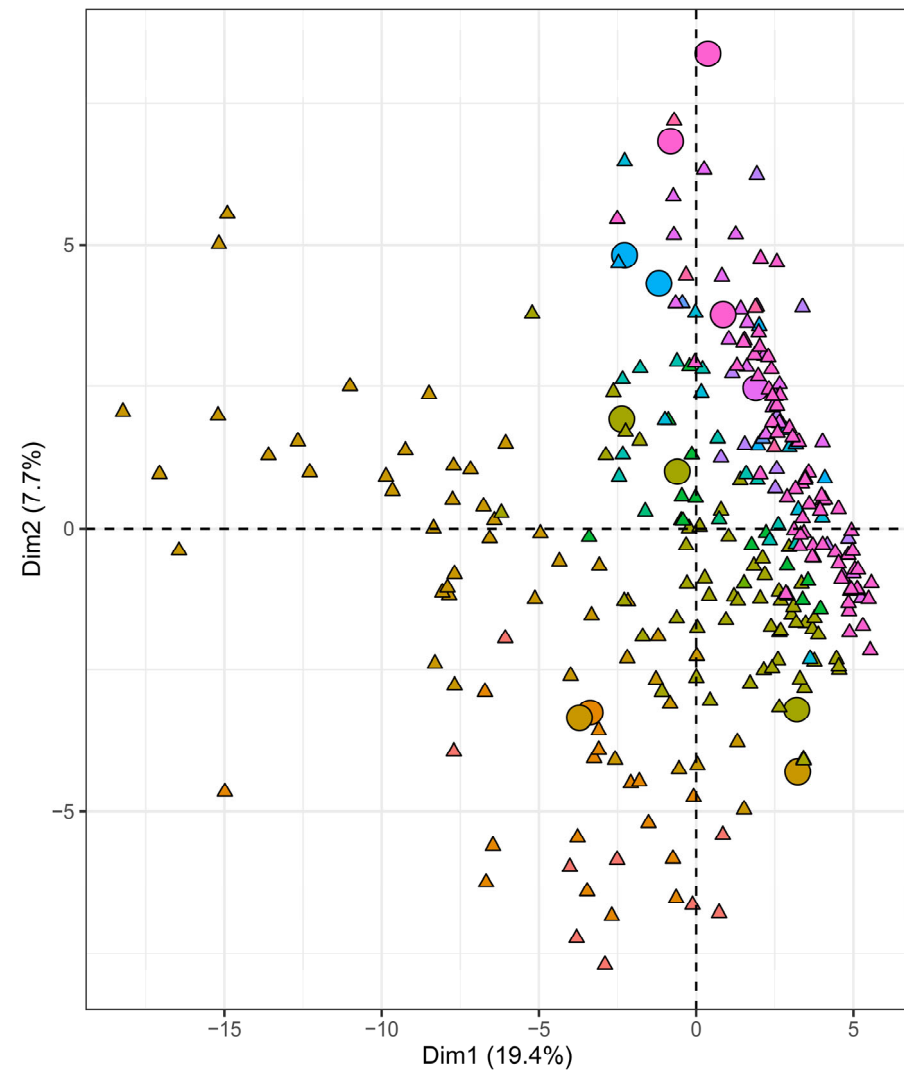
Resource Conservative
“Slow”



Leaf Secondary Metabolism (nonvolatile – HPLC)

All 106 compounds found
2 PCs = 27% of variation

32 metabolites found
across all genotypes
2 PCs = 51% of variation

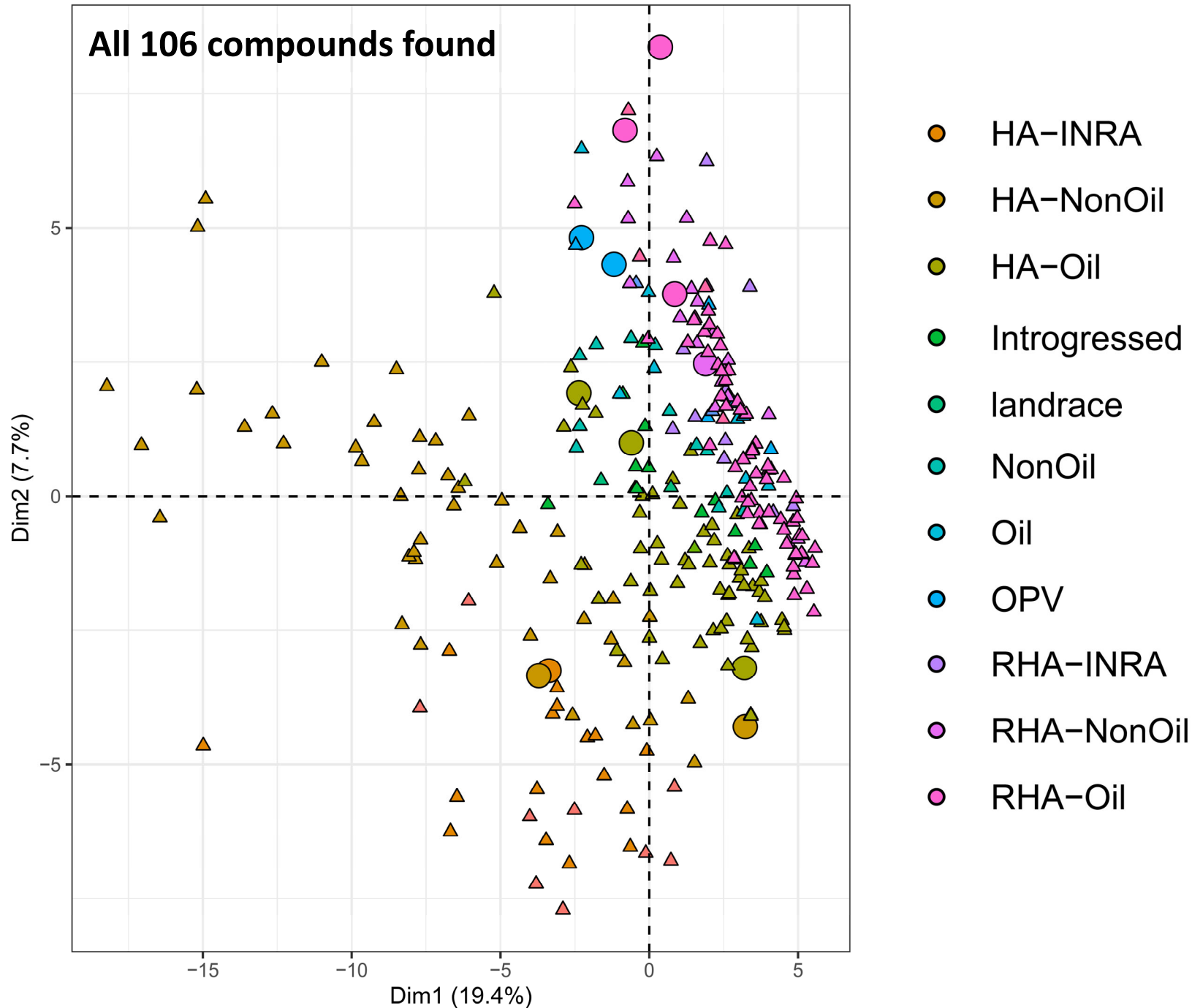


- HA-INRA
- HA-NonOil
- HA-Oil
- Introgressed
- landrace
- NonOil
- Oil
- OPV
- RHA-INRA
- RHA-NonOil
- RHA-Oil

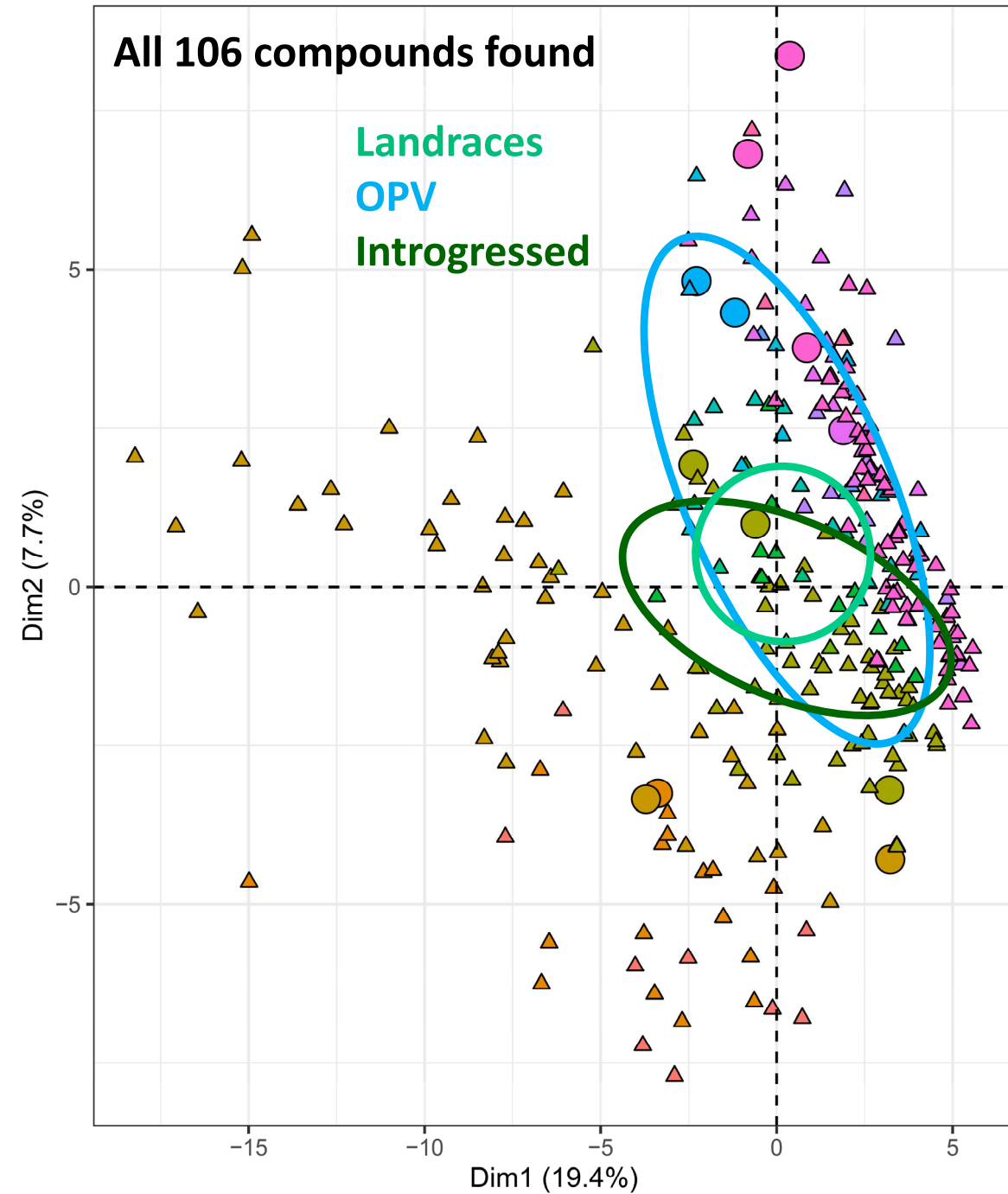
Core 12

- CORE
- △ NON

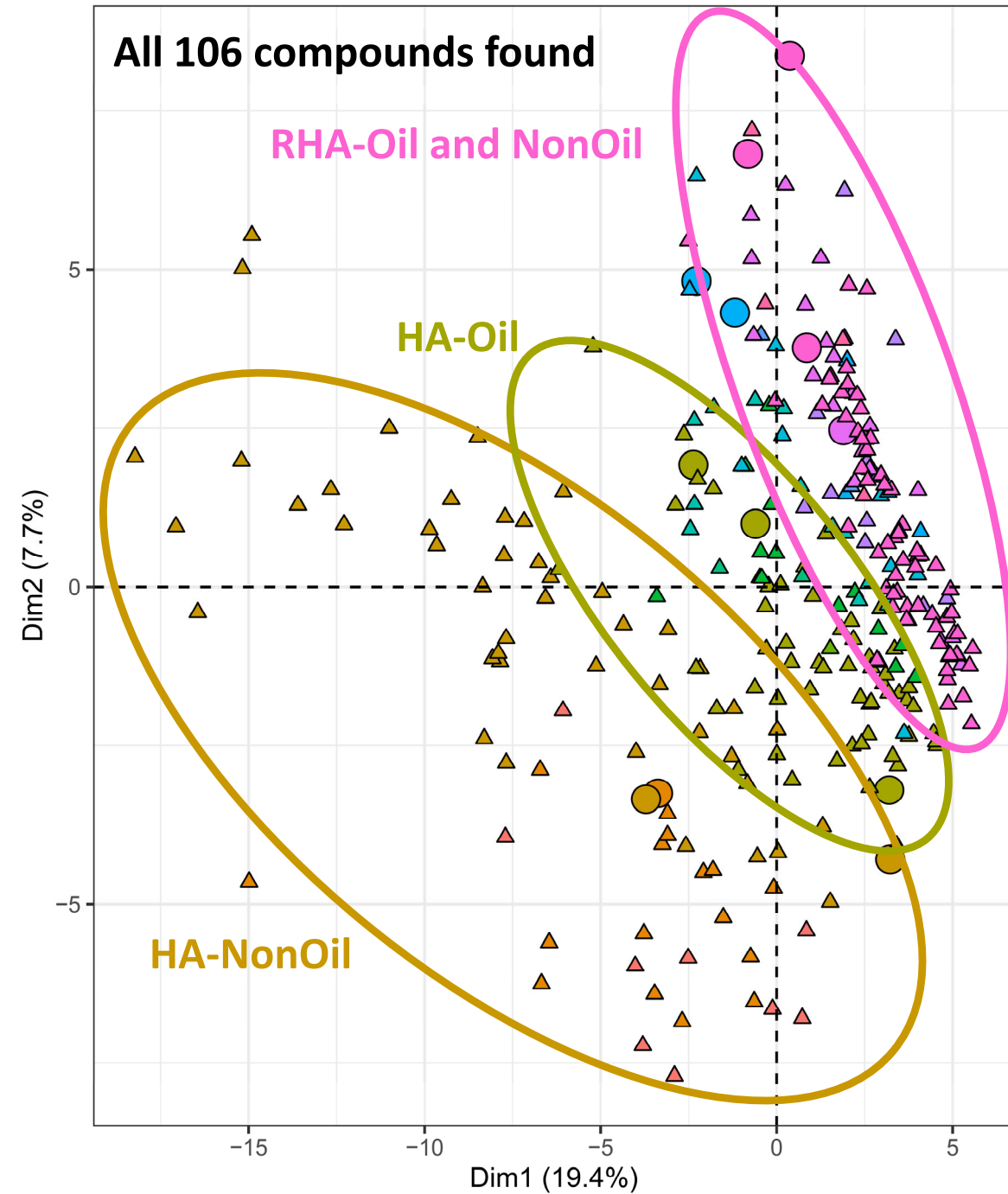
Leaf Secondary Metabolism (nonvolatile – HPLC)



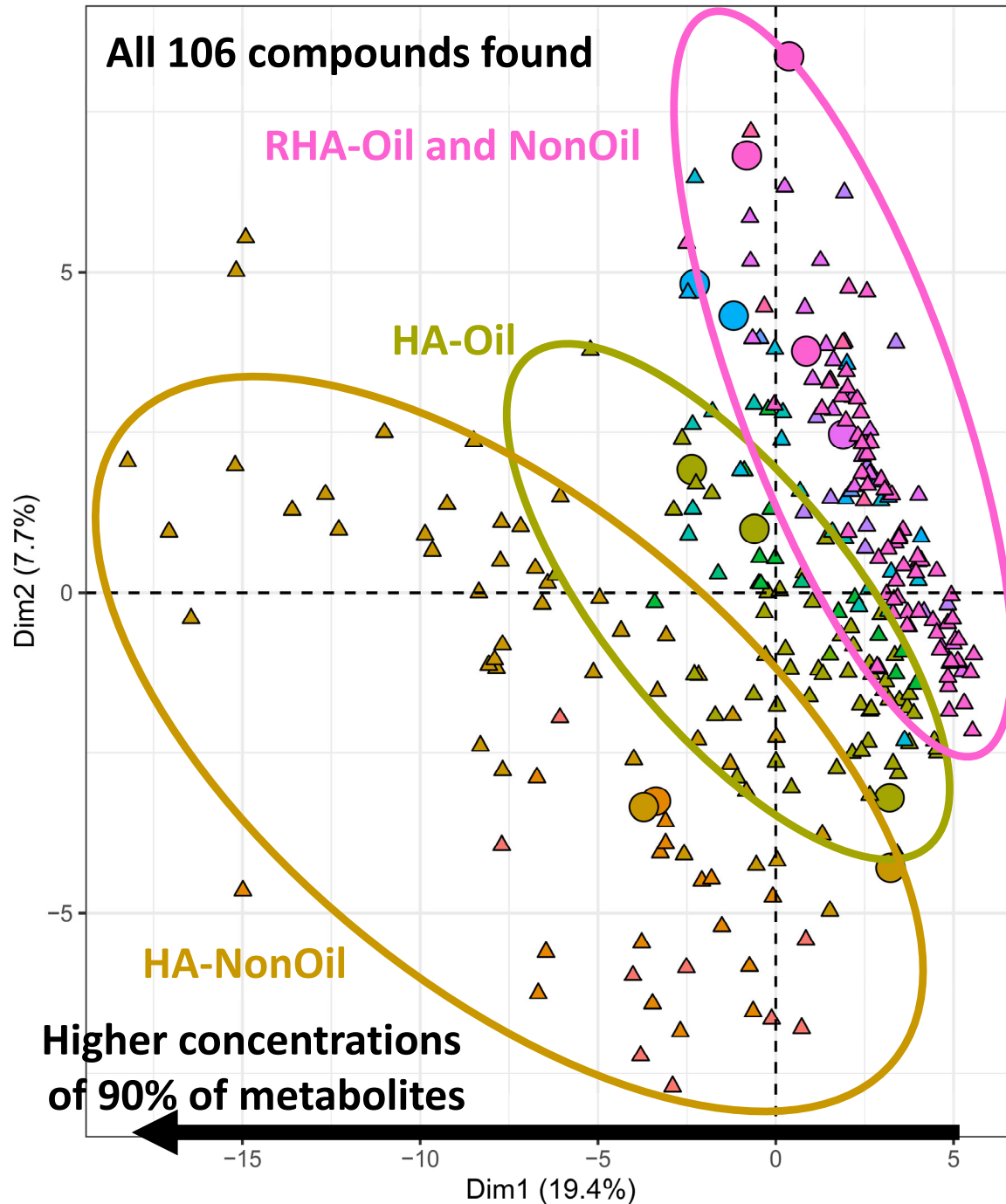
Leaf Secondary Metabolism (nonvolatile – HPLC)



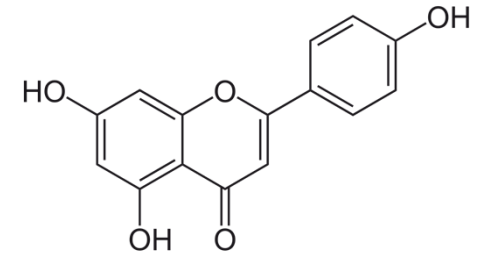
Leaf Secondary Metabolism (nonvolatile – HPLC)



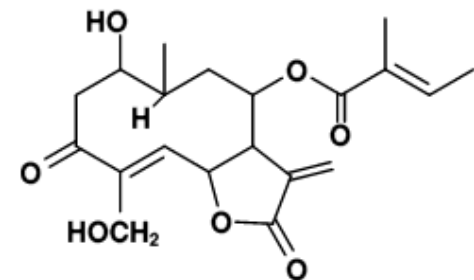
Leaf Secondary Metabolism (nonvolatile – HPLC)



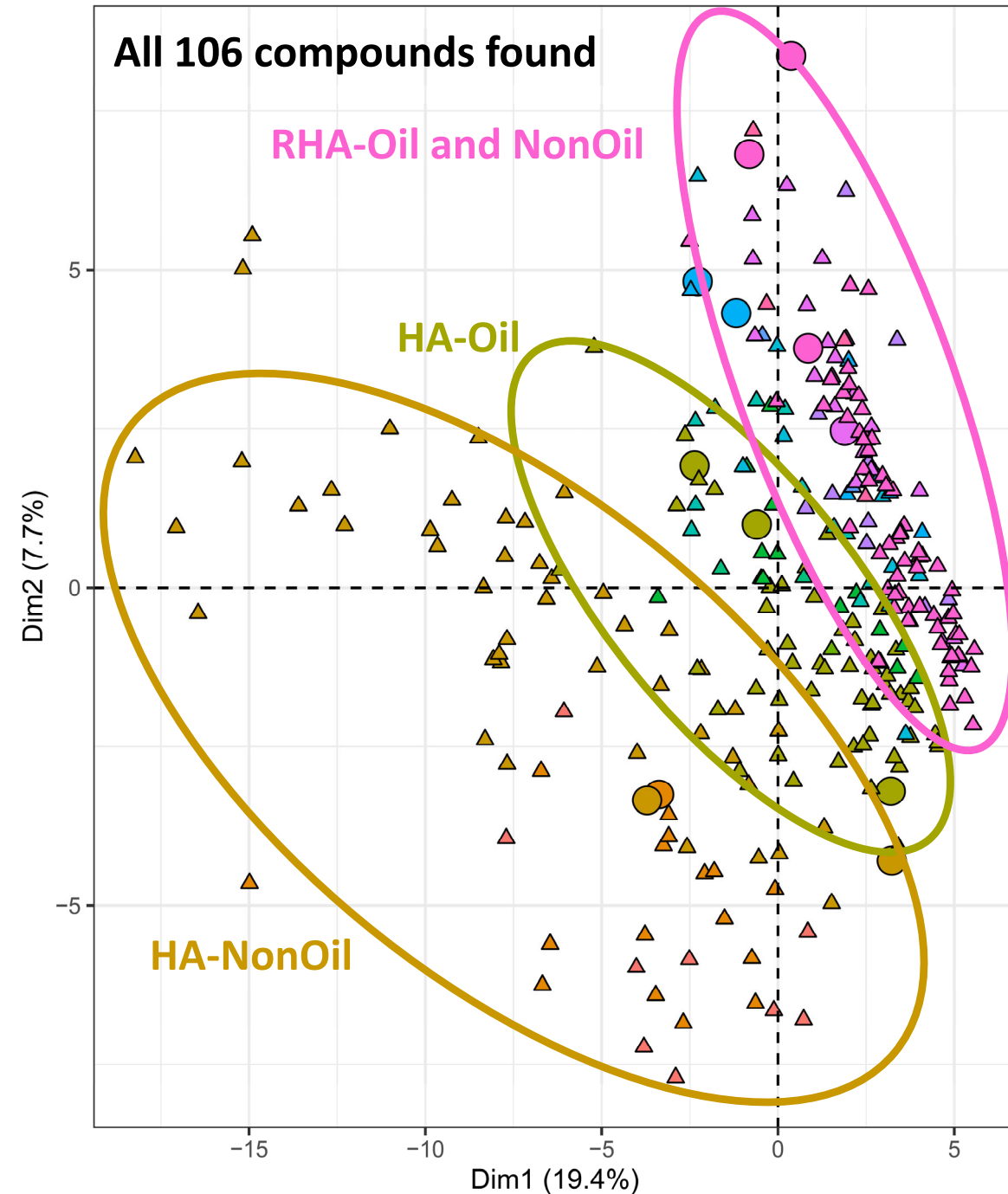
Higher relative concentrations of flavonoids



Higher relative concentrations of sesquiterpene lactones



Leaf Secondary Metabolism (nonvolatile – HPLC)



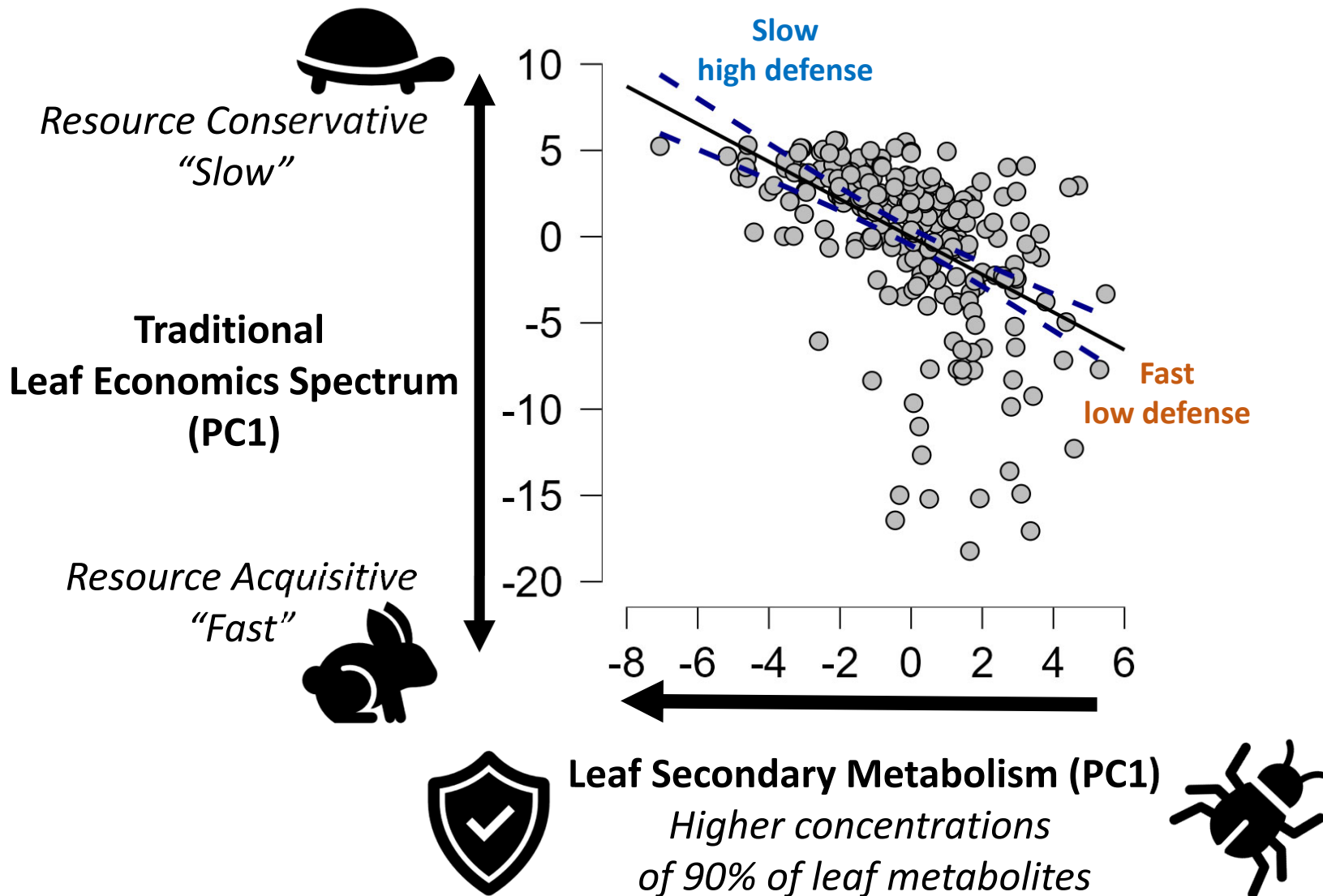
HA-maintainer lines have on average higher concentrations of nonvolatile secondary metabolites than RHA lines – effect size $d=0.24$ ($BF>1000$)

NonOil varieties have on average higher concentrations of nonvolatile secondary metabolites than Oilseed lines – effect size $d=0.23$ ($BF>1000$)

RHA-restorer lines have on average higher flavonoids and HA lines have higher SQTLS – effect size $d=0.21$ ($BF>1000$)

Leaf Ecophysiology + Secondary Metabolism

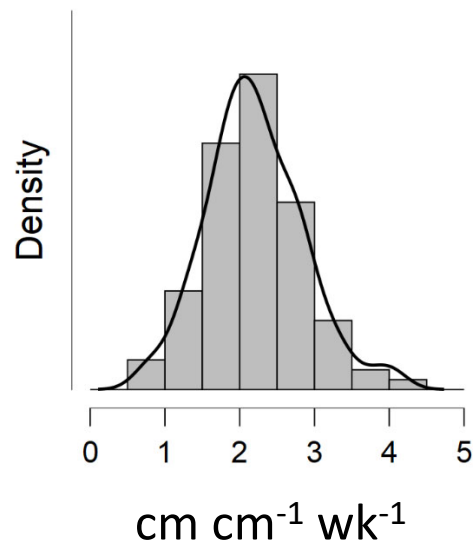
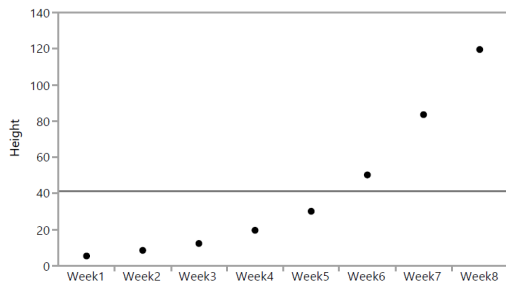
Negative correlation between leaf economic strategy and overall secondary metabolite concentrations ($R^2=0.25$) among lines



Height-based relative growth rate (RGR)

SAM panel grown in a randomized complete block design across two high tunnels under uniform high-resource conditions.

Height assessed weekly for 8 weeks. From this, relative growth rate during peak growth (weeks 3-6) was calculated.



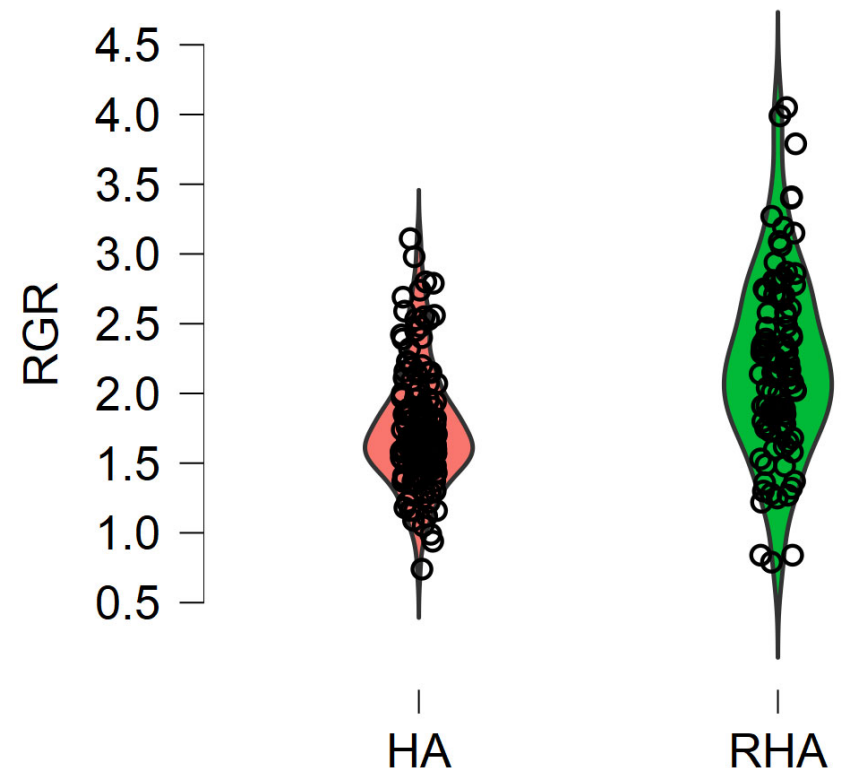
We found a large difference in average Relative Growth Rate between **RHA-restorer** and **HA-maintainer** breeding groups, with RHA having ~30% faster average inherent growth rate than HA.

RHA>HA, effect size $d=0.79$ (BF>1000)

RHA 2.2 cm cm⁻¹ wk⁻¹ [2.1-2.3] 95% Cr.I.

HA 1.7 cm cm⁻¹ wk⁻¹ [1.7-1.8] 95% Cr.I.

No support for overall differences in RGR between Oil and NonOil lines.



So overall...

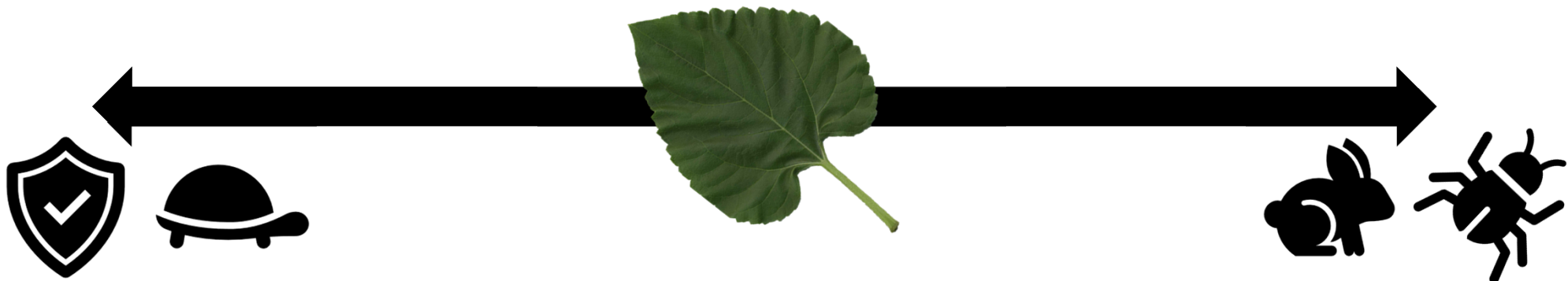
HA-maintainer and **RHA-restorer** lines have diverged in leaf ecophysiology, plant relative growth rate, and *especially* constitutive nonvolatile leaf secondary metabolism.

HA – maintainer

more resource-conservative leaves
lower RGR
higher conc. of metabolites
more SQTs relative to flavonoids

RHA – restorer

more resource-acquisitive leaves
higher RGR
lower conc. of metabolites
more flavonoids relative to SQTs



But *WHY?*

Two potential hypotheses:

(1) **HA-maintainer** lines are more heavily used in breeding efforts for specific target traits, given the ease of performing experimental crosses using male-sterile lines. This, coupled with limited mixing between RHA and HA breeding groups, may explain groups diverging over time and HA diverging farther from OPVs/landraces.

(2) Shifts in breeding goals over time from yield maximization to yield regularity? **RHA-restorer** 'faster' leaf ecophysiology, faster growth, and lower metabolite content may reflect older goals of maximizing total yield, while **HA-maintainer** 'slower' leaf ecophysiology, slower growth, and higher metabolite content may reflect more recent goals of improved yield regularity.

Strong parallels to sunflower wild relatives!

Across wild *Helianthus*, we see very similar axes of trait divergence.

Resource conservative leaves = lower RGR = higher constitutive chemical defense. This suggests that similar constraints on plant form and function exist under artificial selection in crop sunflower.



Next steps – inducible secondary metabolism!

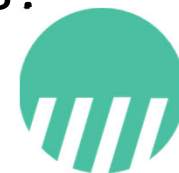
This work focused on constitutive secondary metabolism.

Moving forward, we are using the SAM panel to map the genetic architecture underlying inducible secondary metabolism under attack by a range of pests and pathogens. We hope to determine:

(1) what inducible chemical defenses are ‘universal’ in crop sunflower, versus divergent within the cultivated germplasm?

(2) what inducible chemical defenses are enemy-specific, versus generally activated under attack by multiple pests/pathogens?

(3) are there novel inducible chemical defenses in wild *Helianthus* that could help contribute to polyresistance against enemies like chewing insects and necrotrophic pathogens?



Acknowledgements

Mason + Goolsby Labs @ UCF
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Laura Marek @ USDA



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UCF



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RESEARCH**



Questions?