

GENOMIC REGIONS ASSOCIATED WITH *DIAPORTHE HELIANTHI* AND *DIAPORTHE GULYAE* RESISTANCE

Renan Guidini¹, Mojtaba Jahani², Loren Rieseberg², Laura Marek³, and
Febina Mathew¹

¹Dept. of Agronomy, Horticulture & Plant Science, South Dakota State University, Brookings, SD

▪ ²Dep. of Botany and Biodiversity Research Center, University of British Columbia, Vancouver, British Columbia, Canada

▪ ³USDA-ARS Plant Introduction Research Unit and Department of Agronomy, Iowa State University, Ames, IA

2022 NSA Research Forum



**SOUTH DAKOTA
STATE UNIVERSITY**
*College of Agriculture, Food
and Environmental Sciences*



PHOMOPSIS STEM CANKER

- Yield losses $\geq 40\%$
(Mathew et al. 2015)
- Caused by several fungi of *Diaporthe* genus
- *D. helianthi* and *D. gulyae* are predominant in the U.S. (Elverson et al. 2020)



RESEARCH JUSTIFICATION

- Phomopsis stem canker resistance is quantitative (Vear et al. 1997; Degener et al. 1999; Viguie et al. 1999)
- Identify genes that breeders can use to develop varieties with resistance to *D. gulyae* and *D. helianthi*
- Association mapping has not been performed for *D. gulyae* and *D. helianthi* resistance



RESEARCH OBJECTIVES

1. Assess the disease response associated with *D. helianthi* and *D. gulyae* in the USDA sunflower collection
2. Conduct genome-wide association mapping to identify genomic regions associated with *D. helianthi* and *D. gulyae*
3. Compare the genomic regions conferring resistance to *D. helianthi* and *D. gulyae* with the study by Pogoda & Hulke (2020)

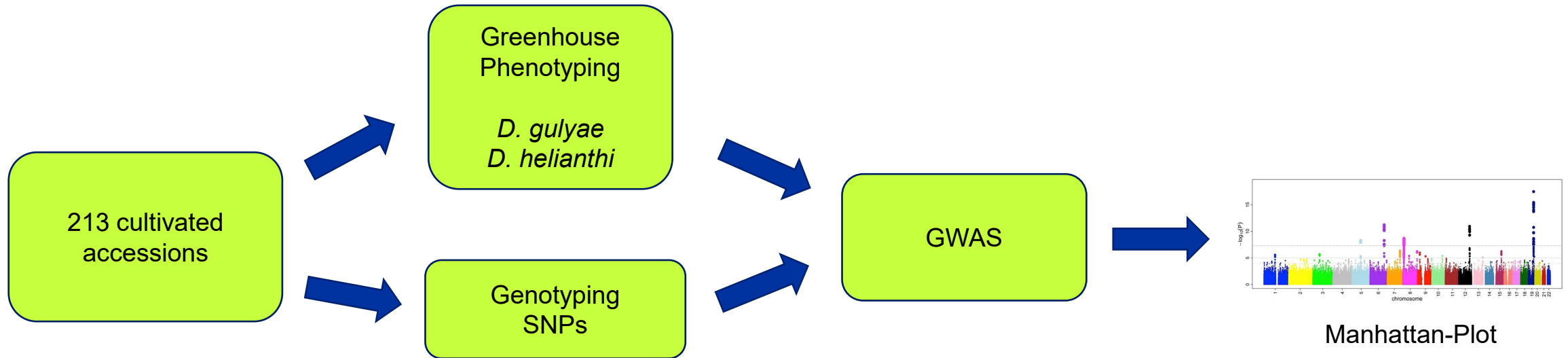


PREVIOUS RESEARCH

- Talukder et al. (2020) identified 15 QTLs that were associated with Phomopsis stem canker resistance
 - 11 chromosomes representing 5.24 to 17.39% of phenotypic variation
 - Recombinant Inbred line population derived from a cross between HA 89 (susceptible) and HA-R3 (resistant)



WORKFLOW



ARTICLES

<https://doi.org/10.1038/s41477-018-0329-0>

nature
plants

Sunflower pan-genome analysis shows that hybridization altered gene content and disease resistance

Sariel Hübner^{1,2,3*}, Natalia Bercovich¹, Marco Todesco¹, Jennifer R. Mandel⁴, Jens Odenheimer⁵, Emanuel Ziegler⁵, Joon S. Lee¹, Gregory J. Baute¹, Gregory L. Owens^{1,6}, Christopher J. Grassa^{1,7}, Daniel P. Ebert^{1,8}, Katherine L. Ostevik^{1,9}, Brook T. Moyers^{1,10}, Sarah Yakimowski¹, Rishi R. Masalia¹¹, Lexuan Gao¹, Irina Čalić¹¹, John E. Bowers¹¹, Nolan C. Kane^{1,12}, Dirk Z. H. Swanevelder^{1,13}, Timo Kubach⁵, Stephane Muñoz¹⁴, Nicolas B. Langlade¹⁴, John M. Burke¹¹ and Loren H. Rieseberg¹



PHENOTYPING

- 213 cultivated accessions from USDA collection
- Consists of open-pollinated varieties, landraces and pre-bred lines (Mandel et al. 2013, 2011)
- Confection inbred 'HA 288' (PI552934) used as the susceptible check (Mathew et al. 2018)



PHENOTYPING

- Screening was performed in the greenhouse at 22-25°C and under a 16 h light/ 8 h dark cycle

- A single isolate of *D. gulyae* and *D. helianthi* used



PHENOTYPING

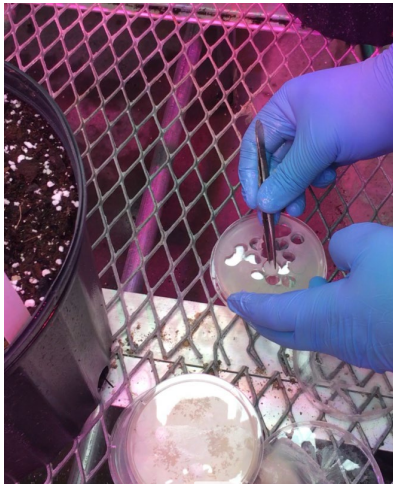


Completely Randomized Design
6 plants per accession
Experiment repeated once

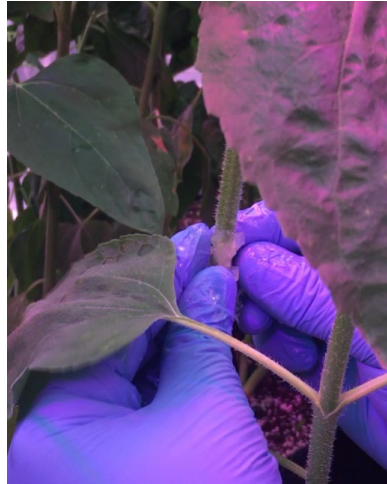


PHENOTYPING

- Inoculation at V4 – V6 (four to six true leaves)
- Mycelial–contact method (Thompson et al. 2011)
- After inoculation, plants misted for 2 min every 2 h for 3 days



6-mm PDA plug



Placed at the third node



Covered with petroleum jelly

PHENOTYPING

- Disease severity evaluated for *D. gulyae* at 14 days after inoculation, and for *D. helianthi* at 30 days
- 0 to 5 disease rating scale (Mathew et al. 2015; Thompson et al. 2011)



0: No discoloration



1: low level discoloration



3: necrotic lesions
2–5 mm, leaf wilting
and twisting



5: very severe
necrosis and lesions,
or plant death

PHENOTYPING DATA ANALYSES

- Disease severity data was analyzed separately for *D. helianthi* and *D. gulyae* using non-parametric statistics

	<i>D. helianthi</i>	<i>D. gulyae</i>
Shapiro-wilk test	$p < 0.0001$	$p < 0.0001$
Levene's test	0.24	0.60



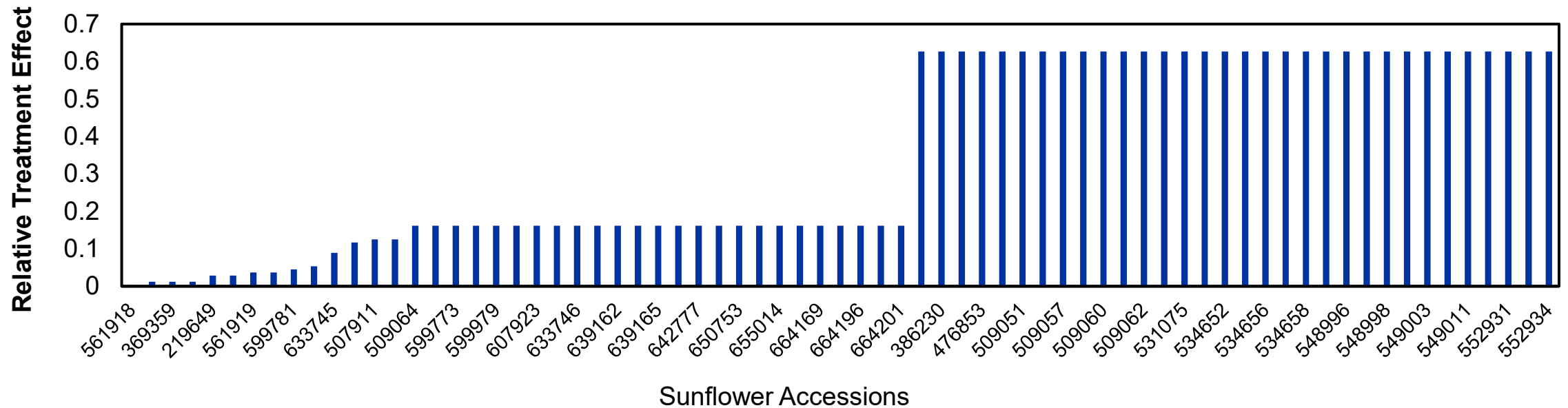
PHENOTYPING DATA ANALYSES

	ANOVA type statistic	Degrees of freedom	<i>p</i> value
<i>D. helianthi</i>	2.16	4.12	<i>p</i> = NS
<i>D. gulyae</i>	10.85	1.54	<i>p</i> < 0.0001

- For *D. helianthi*, since *p*-value was not significant, disease severity associated with the accessions were not compared
- For *D. gulyae*, disease severity was expressed as relative treatment effects and compared using 95% confidence intervals



RESULTS - *D. gulyae*



- 39 accessions had significantly lower RTE compared to HA 288 based on 95% confidence intervals



RESULTS



RESULTS

- 213 accessions were validated for their resistance to Phomopsis stem canker in a field with disease history (natural inoculum)

- PI 509060, PI 561918, PI 599782, and PI 633745 had significantly lower RTE compared to HA 288 in the field and greenhouse



GENOTYPING

- Genomic DNA extracted from each accession using CTAB protocol (Todesco et al. 2020)
- Whole-genome shotgun Illumina libraries prepared using TruSeq protocol (Rowan et al. 2015; Rohland and Reich 2012)
- Sequencing conducted on the HiSeq platform with 150-bp paired-end reads



GENOTYPING

- Sequences were trimmed (Trimmomatic v.036) and aligned to the *Helianthus annuus* XRQv1 genome (3.6 Gbp) (NextGenMap (5.3) (Bolger et al. 2014)
- Variants calling followed the Genome Analysis Toolkit (Poplin et al. 2017)
- 3,647,583 biallelic SNPs with minor allele frequency > 0.03

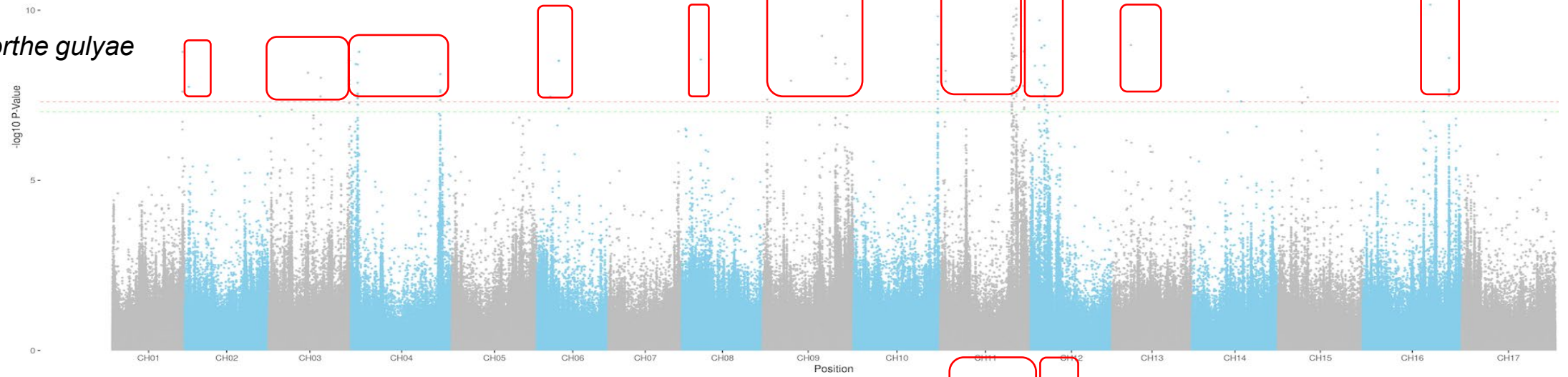


GENOME-WIDE ASSOCIATION

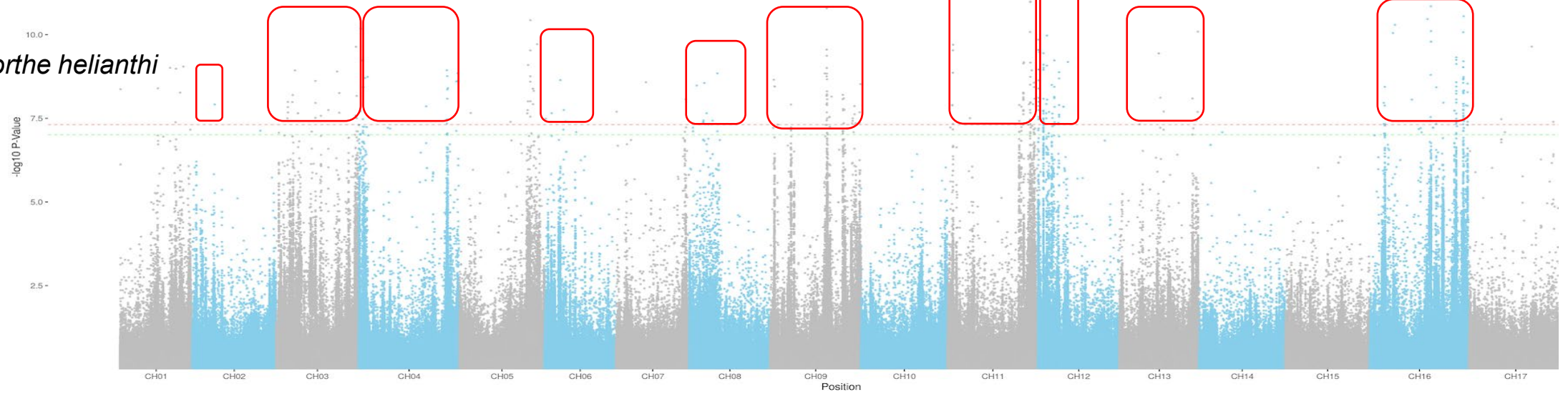
- EMMAX software with genotype as a fixed effect (Kang et al. 2010)
- To control false positives, a mixed model was employed with K- and P-matrices as covariates (Kang et al. 2010)
- To identify significant associations, a correction method for multiple testing was implemented by $-\log_{10}(0.05 \times M_{\text{eff}}^{-1})$ (Gao 2011)




Diaporthe gulyae



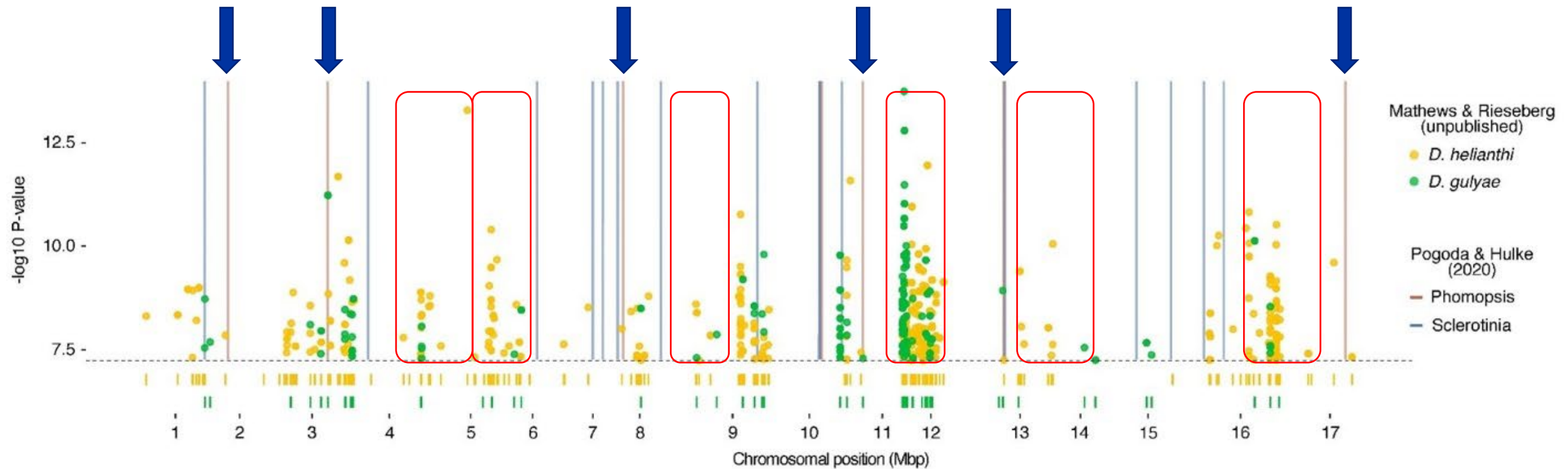
Diaporthe helianthi



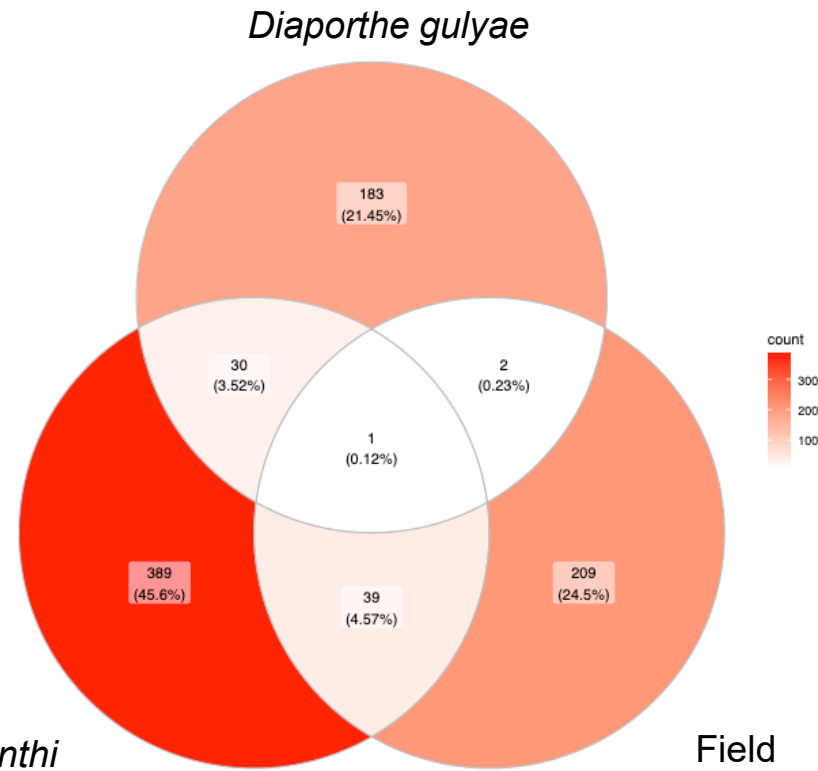
Genetic loci underlying quantitative resistance to necrotrophic pathogens *Sclerotinia* and *Diaporthe* (*Phomopsis*), and correlated resistance to both pathogens

Cloe S. Pogoda¹ · Stephan Reinert¹ · Zahirul I. Talukder² · Ziv Attia¹ · Erin C. E. Collier-zans¹ · Thomas J. Gulya³ · Nolan C. Kane¹ · Brent S. Hulke³ 

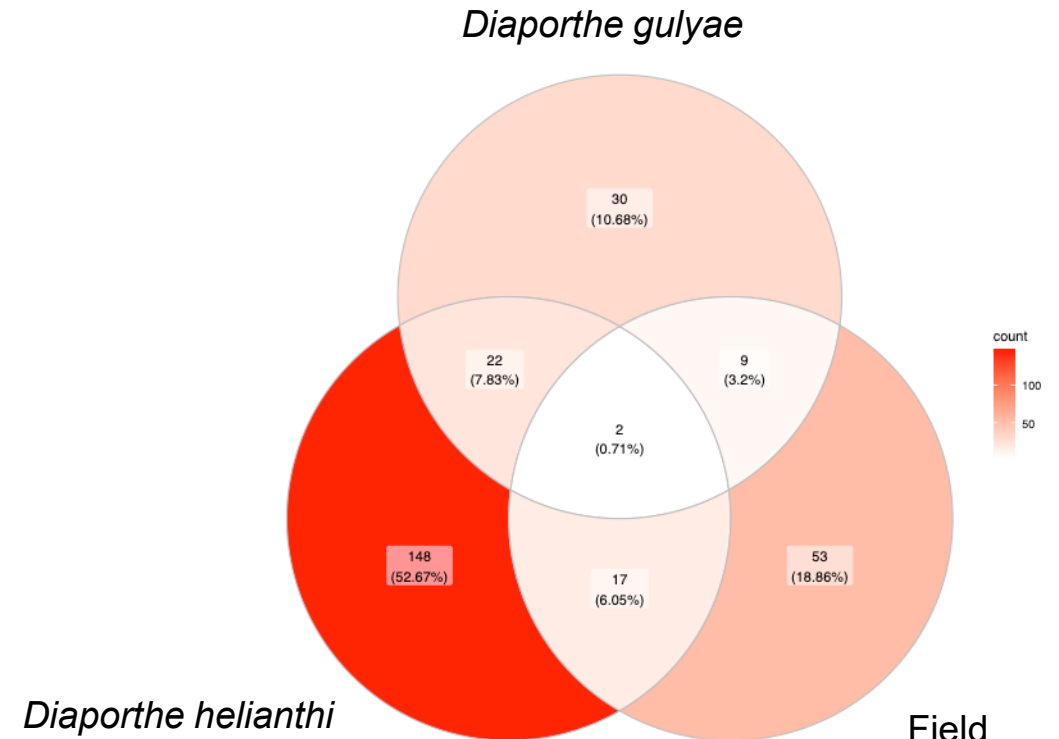
Received: 19 June 2020 / Accepted: 18 September 2020 / Published online: 27 October 2020
© Springer-Verlag GmbH Germany, part of Springer Nature 2020



SNPs overlap



Genes overlap



- 30 SNPs associated with resistance to *D. gulyae* and *D. helianthi*

- 22 genes associated with resistance to *D. gulyae* and *D. helianthi*

SUMMARY

- This study provides information for sunflower improvement through identification of parental materials which can be exploited in breeding programs.

- 39 accessions had significantly lower RTE, among which PI 509060, PI 561918, PI 599782, and PI 633745 had significantly lower RTE compared to HA 288 in the field



SUMMARY

- 10 chromosomes (2, 3, 4, 6, 8, 9, 11, 12, 13, and 16) were associated with resistance for *D. gulyae* and *D. helianthi*
- 6 chromosomes (2, 3, 8, 11, 13, and 17) were common between the study by Pogoda and Hulke (2020) and this study



SUMMARY

- 30 SNPs and 22 genes were associated with resistance to *D. gulyae* and *D. helianthi*

- SNPs with the highest effect size were identified on chromosomes 4 and 11



IMPLICATION

- Markers flanking the *D. gulyae* and *D. helianthi* resistance will facilitate marker-assisted selection in breeding

- Following breeding and development, disease resistant varieties can be incorporated into an IPM program for Phomopsis stem canker



FUTURE WORK

- Identify and validate candidate genes associated with resistance to *D. gulyae* and *D. helianthi* around the GWAS-identified loci

- Determine the expression level changes in genes located in GWAS-identified loci through RNA-seq analysis



LITERATURE CITED

- Bolger, A. M., Lohse, M., and Usadel, B. 2014. Trimmomatic: a flexible trimer for Illumina sequence data. *Bioinformatics* 30: 2114-2120.
- Degener, J., Melchinger, A. E., and Hahn, V. 1999. Inheritance of resistance to Phomopsis in sunflower study of leaf and stem resistance after artificial and natural infection. *Helia* 22: 105–116.
- Elverson, T., Kontz, B., Markell, S., Harveson, R.M., and Mathew, F. 2020. Quantitative PCR assays developed for *Diaporthe helianthi* and *Diaporthe gulyae* for Phomopsis stem canker diagnosis and germplasm screening in sunflower (*Helianthus annuus*). *Plant Dis.* 104:793-800.
- Gao, X. Y. 2011. Multiple testing corrections for imputed SNPs. *Genetic Epidemiology* 35: 154–158
- Kang, H. M., Sul, J. H., Service, S. K., Zaitlen, N. A., Kong, S. Y., Freimer, N. B., Sabatti, C., and Eskin, E. 2010. Variance component model to account for sample structure in genome-wide association studies. *Nature Genetics* 42: 348–354.
- Mandel, J.R., Dechaine, J.M., Marek, L.F., and Burke, J.M. 2011. Genetic diversity and population structure in cultivated sunflower and a comparison to its wild progenitor, *Helianthus annuus* L. *Theor. Appl. Genet.* 123:693–704.



LITERATURE CITED

- Mandel J.R, Nambeesan S., Bowers J. E., Marek L. F., Ebert D., Rieseberg L. H., Knapp S. J., and Burke J., M. 2013. Association mapping and the genomic consequences of selection in sunflower. PLoS Genetics 9: e1003378.
- Mathew, F. M., Alananbeh, K. M., Jordahl, J. G., Meyer, S. M., Castlebury, L. A., Gulya, T. J., and Markell, S. G. 2015. Phomopsis stem canker: A reemerging threat to sunflower (*Helianthus annuus*) in the United States. Phytopathology 105:990-997.
- Mathew, F., Olson, T., Marek, L., Gulya, T., and Markell, S. 2018. Identification of sunflower (*Helianthus annuus*) accessions resistant to *Diaporthe helianthi* and *Diaporthe gulyae*. Plant Health Prog. 19: 97–102.
- Poplin, R. et al. 2017. Scaling accurate genetic variant discovery to tens of thousands of samples. Preprint at <https://www.biorxiv.org/content/10.1101/201178v3>
- Rowan, B. A., Patel, V., Weigel, D., and Schneeberger, K. 2015. Rapid and inexpensive whole-genome genotyping-by-sequencing for crossover localization and fine-scale genetic mapping. G3 5: 385–398.
- Rohland, N., and Reich, D. 2012. Cost-effective, high-throughput DNA sequencing libraries for multiplexed target capture. Genome Res. 22: 939–946.



LITERATURE CITED

- Talukder, I. Z., Underwood, W., Ma, G., Seiler, J. G., Misar, G. C., Cai, X., and Qi, L. 2020. Genetic dissection of *Phomopsis* stem canker resistance in cultivated sunflower using high density SNP linkage map. *Int. J. Mol. Sci.* 1497; doi:103390/ijms21041497
- Thompson, S.M., Tan, Y.P., Young, A.J., Neate, S.M., Aitken, E.A.B., and Shivas, R.G. 2011. Stem cankers on sunflower (*Helianthus annuus*) in Australia reveal a complex of pathogenic *Diaporthe* (*Phomopsis*) species. *Persoonia* 27:80-89.
- Todesco, M., Owens, G.L., Bercovich, N. *et al.* 2020. Massive haplotypes underlie ecotypic differentiation in sunflowers. *Nature* 584: 602–607.
- Vear, F., Garreyn, M., and Tourvieille de Labrouhe, D. 1997. Inheritance of resistance to *Phomopsis* (*Diaporthe helianthi*) in sunflower. *Plant Breed.* 116: 277–281. <http://dx.doi.org/10.1111/j.1439-0523.1997.tb00996.x>
- Viguié, A., Vear, F., and Tourvieille de Labrouhe, D. 1999. Interaction between French isolates of *Phomopsis/Diaporthe helianthi* Munt.-Cvet. et al. and sunflower (*Helianthus annuus* L.) genotypes. *Eur. J. Plant Pathol.* 105: 693–702.



THANK YOU!

My lab:

Nathan Braun

Brian Kontz

Ruchika Kashyap

Nabin Dangal

Karthika Mohan

Bijula Sureshbabu

Ellen Tuschen

Samantha Schryver

Dr. Shyam Solanki

renan.guidini@sdstate.edu



**SOUTH DAKOTA
STATE UNIVERSITY**
*College of Agriculture, Food
and Environmental Sciences*

